Lambda-Cyhalothrin Criteria Derivation DRAFT

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1. Introduction

A new methodology for deriving freshwater water quality criteria for the protection of aquatic life was developed by the University of California, Davis (TenBrook et al. 2009a). The need for a new methodology was identified by the California Central Valley Regional Water Quality Control Board (CVRWQCB 2006) and findings from a review of existing methodologies (TenBrook & Tjeerdema 2006, TenBrook et al. 2009b). This new methodology is currently being used to derive aquatic life criteria for several pesticides of particular concern in the Sacramento River and San Joaquin River watersheds. The methodology report contains an introduction (Chapter 1); the rationale of the selection of specific methods (Chapter 2); detailed procedure for criteria derivation (Chapter 3); and a chlorpyrifos criteria report (Chapter 4). This criteria report for lambda-cyhalothrin describes, section by section, the procedures used. Also included are references to specific sections of the methodology procedure detailed in Chapter 3 of the report so that the reader can refer to the report for further details (TenBrook et al. 2009a).

2. Basic information

Chemical: Lambda-cyhalothrin (Fig. 1)

CAS: $[1\alpha(S^*), 3\alpha(Z)]$ -(±)-cyano-(3-phenoxyphenyl)methyl 3-(2-chloro-3,3,3-trifluoro-1propenyl)-2,2-dimethylcyclopropanecarboxylate

IUPAC: (S)- α -cyano-3-phenoxybenzyl (Z)-(1R,3R)-3-(2-chloro-3,3,3-trifluoropropenyl)-2,2-dimethylcyclopropanecarboxylate

Chemical Formula: C₂₃H₁₉ClF₃NO₃

CAS Number: 91465-08-6 CA DPR Chem Code: 2297 USEPA PC Code: 128897

Trade names: Warrior, Phoenix, SFK, Charge, Excaliber, Grenade, Hallmark, Icon, Karate, Matador, OMS 0321, PP321, Saber, Samurai and Sentinel (ExToxNet 1995,

Tomlin 2003).

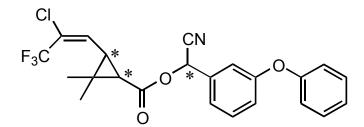


Figure 1. Structure of lambda-cyhalothrin, asterisks indicate stereocenters.

3. Physical-chemical data

Molecular Weight

449.850 Mackay et al. 2006

Composition

Equal quantities of (S)- α -cyano-3-phenoxybenzyl(Z)-(1R,3R)-3-(2-chloro-3,3,3-trifluoropropanyl)-2,2-dimethylcyclopropanecarboxylate and (S)- α -cyano-3-phenoxybenzyl(Z)-(1S,3S)-3-(2-chloro-3,3,3-trifluoropropanyl)-2,2-dimethylcyclopropanecarboxylate (Tomlin 2003). Technical grade is ~81% pure (Tomlin 2003).

Density

1.33 g/mL at 25°C Tomlin 2003, Mackay *et al.* 2006

Water Solubility

 0.005 mg/L at 20°C (mean, n=12)
 Laskowski 2002

 0.005 mg/L at 20°C (pH 6.5)
 Mackay et al. 2006

 0.005 mg/L at 20°C (pH 6.5, purified water)
 Tomlin 2003, Mackay et al. 2006

 0.004 mg/L (pH 5.0, buffered water)
 Mackay et al. 2006

Melting Point

49.2°C Tomlin 2003 Technical: 47.5-48.5°C Tomlin 2003

Vapor Pressure

 1.56 x 10⁻⁹ mm Hg at 20°C
 Laskowski 2002

 2.0 x 10⁻⁷ Pa at 20°C
 Mackay et al. 2006

 2.0 x 10⁻⁷ Pa at 20°C (estimated)
 Tomlin 2003, Mackay et al. 2006

 2.0 x 10⁻⁴ Pa at 60°C (interpolated)
 Tomlin 2003, Mackay et al. 2006

 7.80 x 10⁻⁶ Pa at 40°C
 Mackay et al. 2006

 19 x 10⁻⁶ Pa at 40°C (measured 40-80°C)
 Mackay et al. 2006

Organic Carbon Sorption Partition Coefficients ($\log K_{oc}$)

5.52 soil and sediment organic matter Tomlin 2003 soil adsorption (mean of 50 experiments) 5.51 Laskowski 2002

GeoMean of $log K_{oc}$ values: 5.52

 $\frac{\text{Henry's constant }(K_{H})}{1.9 \text{ x } 10^{\text{-}7} \text{ atm m}^{3} \text{ mol}^{\text{-}1} \text{ at } 20^{\circ}\text{C}} \\ 2 \text{ x } 10^{\text{-}2} \text{ Pa m}^{3} \text{ mol}^{\text{-}1}$ Laskowski 2002 Tomlin 2003

 $Log K_{ow}$

7.0 at 20°C Mackay et al. 2006, Laskowski 2002, Tomlin 2003

6.1 (calculated from molecular structure) Laskowski 2002

 pK_a

> 9 (hydrolysis prevents measurement) Tomlin 2003

Table 1. Bioconcentration factors (BCF) for cyhalothrin and lambda-cyhalothrin; FT:

flow-through.

Species	BCF	Exposure	Reference
Cyprinus carpio	2240	FT	Yamauchi et al. 1984
(whole fish)			(cyhalothrin)
Cyprinus carpio	7340	FT	Yamauchi et al. 1984
(viscera)			(cyhalothrin)
Cyprinus carpio	850	FT	Yamauchi et al. 1984
(muscle)			(cyhalothrin)
Chironomus	2000	Water only	Hamer <i>et al</i> . 1999
riparius			
Chironomus	2300 (mean)	Water-	Hamer <i>et al</i> . 1999
riparius		sediment	
Daphnia magna	194	Water-	Hamer & Hill 1985 (cyhalothrin)
		sediment	
Ictalurus	19	Water-	Hamer & Hill 1985 (cyhalothrin)
punctatus		sediment	
(whole fish)			
Ictalurus	7	Water-	Hamer & Hill 1985 (cyhalothrin)
punctatus		sediment	
(muscle)			
Ictalurus	66	Water-	Hamer & Hill 1985 (cyhalothrin)
punctatus		sediment	
(viscera)			

Table 2. Lambda-cyhalothrin hydrolysis, photolysis, and biodegradation.

	Half- life (d)	Water	Temp (°C)	pН	Reference
Hydrolysis	Stable (0 d)	Sterile,	25	5	Laskowski
		buffered			2002
	Stable (0 d)	Sterile,	25	7	Laskowski
		buffered			2002
	8.66	Sterile,	25	9	Laskowski
		buffered			2002
Aqueous	24.5	NR	25	5	Laskowski
Photolysis					2002
Biodegradation	21.9	Natural water	20	NR	Laskowski
(aerobic)					2002

4. Human and wildlife dietary values

There are no FDA action levels for lambda-cyhalothrin (USFDA 2000). There are no food tolerances for fish, but there are food tolerances for the meat of hogs at 0.1 ppm (USEPA 2007).

Wildlife LC₅₀s (dietary) for animals with significant food sources in water

The dietary LC_{50} for 8-d old mallard ducks was determined to be 3948 ppm (Roberts *et al.* 1985). In another acute oral toxicity test with mallard duck a dietary LD_{50} could not be determined because there were no effects observed at any of the concentrations; the highest concentration tested was 3950 mg/kg of feed (Roberts & Fairley 1984).

Wildlife dietary NOECs for animals with significant food sources in water

A dietary NOEL of 30 mg/kg of feed for mallard ducks was determined over a 20 week period (Beavers *et al.* 1990). A LOEL could not be determined in this study because no significant effects were observed at any concentration tested. The highest concentration of lambda-cyhalothrin in mallard feed was 30 mg/kg, which was reported as the NOEL for the study.

Lambda-cyhalothrin did not bioaccumulate in mallard ducks over a 28-d exposure given by oral gavage (Knight & Leahey 1984).

5. Ecotoxicity data

Approximately 65 original studies of the effects of lambda-cyhalothrin on aquatic life were identified and reviewed. In the review process, many parameters are rated for documentation and acceptability for each study, including, but not limited to: organism source and care, control description and response, chemical purity, concentrations tested, water quality conditions, and statistical methods (see Tables 3.6, 3.7, 3.8 in TenBrook *et*

al. 2009a). Single-species effects studies that were rated relevant (R) or less relevant (L) according to the method were summarized in the data summary sheets. Information in these summaries was used to evaluate each study for reliability using the rating systems described in the methodology (section 3-2.2, TenBrook et al. 2009a). Copies of completed summaries for all studies are included in Appendix A of this report. Lambda-cyhalothrin studies deemed irrelevant from an initial screening were not summarized (e.g., studies involving rodents or in vitro exposures). All data rated as acceptable or supplemental for criteria derivation are summarized in Tables 3 - 9 found at the end of this report.

Using the data evaluation criteria (section 3-2.2, TenBrook *et al.* 2009a), 20 acute toxicity studies, yielding 66 toxicity values, were judged reliable and relevant (RR) for criteria derivation (Tables 3 and 4). Three chronic toxicity studies, yielding twelve toxicity values, were judged reliable and relevant (RR) for criteria derivation (Tables 6 and 7). Six acute and three chronic studies were rated RL, LL, or LR and were used as supplemental information for evaluation of the derived criteria in section 12 (Tables 5 and 9).

Eleven mesocosm, microcosm and ecosystem (field and laboratory) studies were identified and reviewed. Eight of these studies were rated R or L and were used as supporting data in section 13 (Table 10). Four studies of lambda-cyhalothrin effects on wildlife were identified and reviewed for consideration of bioaccumulation in section 15.

6. Data reduction

Multiple toxicity values for lambda-cyhalothrin for the same species were reduced into one species mean acute toxicity value according to procedures described in the methodology (section 3-2.4, TenBrook *et al.* 2009a). Acceptable acute and chronic data that were excluded, and the reasons for their exclusion, are shown in Tables 4 and 7, respectively. Reasons for exclusion of data included: flow-through tests are preferred over static tests, a test with a more sensitive life-stage of the same species was available, more sensitive endpoints were available for the same test, and more appropriate or more sensitive test durations were available for the same test. The final acute and chronic data sets are shown in Tables 3 and 6, respectively.

7. Acute criterion calculation

At least five acceptable acute toxicity values were available and fulfilled the five taxa requirements of the species sensitivity distribution (SSD) procedure (section 3-3.1, TenBrook *et al.* 2009a). The five taxa requirements are a warm water fish, a cold water fish, a planktonic crustacean, a benthic crustacean, and an insect. The Burr Type III SSD procedure was used for the acute criterion calculation because more than eight acceptable acute toxicity values were available in the lambda-cyhalothrin data set as seen in Table 3 (section 3-3.2.2, TenBrook *et al.* 2009a). The Burr Type III SSD procedure was used to derive 5th percentile values (median and 95% confidence limit), as well as 1st percentile values (median value only, as the software could not provide a 95% confidence limit for

the 1st percentile). Comparing the 95% confidence limit to the acute criteria, it can be seen that there is uncertainty in the first significant figure, thus the final criterion will be reported with one significant digit.

Burr III distribution

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Fit parameters: b=0.232356; c=1.100750; k=0.596085 (likelihood=-4.987264)
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5^{th} percentile, 50% confidence limit: 0.002432 µg/L 5^{th} percentile, 95% confidence limit: 0.000501 µg/L 1^{st} percentile, 50% confidence limit: 0.000208 µg/L
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Recommended acute value = $0.002432 \mu g/L$ (median 5th percentile value)

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Acute criterion = acute value \div 2
= 0.002432 \div 2 = 0.001216 \mug/L
Acute criterion = 0.001 \mug/L
= 1 ng/L
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Acute values were plotted in a histogram (Figure 2). The data do not appear to be bimodal. The fit of the Burr III distribution from the BurrliOZ software is shown in Figure 3. This distribution provided a satisfactory fit (see Appendix B) according to the fit test described in section 3-3.2.4 of TenBrook *et al.* (2009a). No significant lack of fit was found ($\chi^2_{2n} = 0.1994$) using a fit test based on cross validation and Fisher's combined test (section 3-3.2.4, TenBrook *et al.* 2009a), indicating that the data set is valid for criteria derivation.

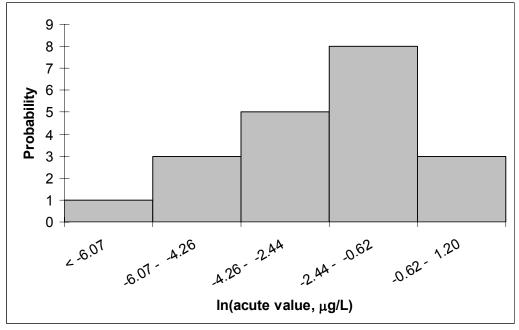


Figure 2. The natural log of the lambda-cyhalothrin species mean acute values were plotted on a histogram to show the general shape of the distribution of the data.

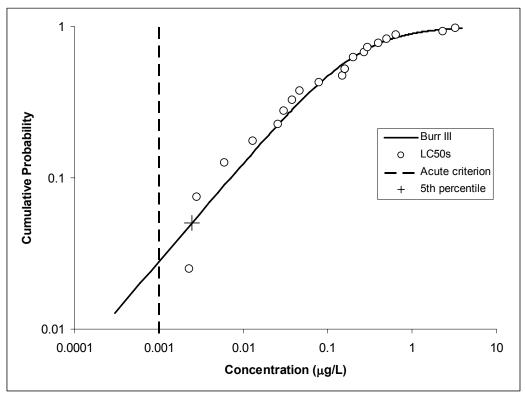


Figure 3. The fit of the Burr III distribution plotted with the acute toxicity values.

8. Chronic criteria calculation

Chronic toxicity values from fewer than five different families were available, thus the acute-to-chronic ratio (ACR) method was used to calculate the chronic criterion (section 3-4.2, TenBrook *et al.* 2009a). Two chronic toxicity values are in the acceptable (rated RR) data set (Table 6) satisfying two of the five taxa requirements (section 3-3.1, TenBrook *et al.* 2009a): warm water fish (*Pimephales promelas*) and planktonic crustacean (*Daphnia magna*).

Two of the chronic toxicity values could be paired with an appropriate corresponding acute toxicity value in order to calculate an ACR, satisfying two of the three family requirements: a fish and an invertebrate (section 3-4.2.1, TenBrook *et al.* 2009a). Data for one additional acutely sensitive species is required to derive an ACR based on measured data. Because there were only acceptable chronic data for two freshwater species, data from a saltwater species was used to fulfill the final requirement because freshwater and saltwater ACRs have been shown to be comparable (section 3-4.2.1, TenBrook *et al.* 2009a). Saltwater data in the supplemental data sets (Tables 5 and 9) contained acute and chronic toxicity values for Sheepshead minnow (*Cyprinodon variegatus*) from the same flow-through study conducted by the same laboratory in the same dilution water, which are appropriate data for ACR derivation (section 3-4.2.1, TenBrook *et al.* 2009a).

The ACRs were calculated for each of the three species by dividing the acute LC₅₀ value by the chronic MATC value. The final multi-species ACR was obtained by calculating the geometric mean of the three ACR values because all species were within a factor of ten and there was not an increasing or decreasing trend in species mean ACR (SMACR) values with the species mean acute values (step 2, section 3-4.2.1, TenBrook *et al.* 2009a). The individual species and final multi-species ACR values generated are shown in Table 8.

The chronic criterion was calculated using the acute median 5th percentile value and the final multi-species ACR value as follows:

Chronic criterion = acute median 5^{th} percentile value ÷ ACR = $0.002432 \mu g/L \div 4.73 = 0.0005144 \mu g/L$ Chronic criterion = $0.001 \mu g/L$ = 1 ng/L

This value is a factor of 2.63 below the lowest acceptable chronic value (MATC) of 0.00263 µg/L for *Daphnia magna* (Table 6).

9. Bioavailability

Although lambda-cyhalothrin and other pyrethroids are not very soluble in water, aquatic organisms are very sensitive to pyrethroids and toxicity does occur. Pyrethroids have been found as the cause of toxicity in surface waters in the California Central Valley. This toxicity is believed to occur primarily from the portion of the compound that is dissolved in the water, not from the compound that is associated with the particulate phase (Amweg *et al.* 2005).

Several studies suggest that the binding of lambda-cyhalothrin and other pyrethroids to suspended solids and dissolved organic matter will make the bound fraction unavailable and thus nontoxic to aquatic organisms. The effects of dissolved organic carbon (DOC) on the acute toxicity of cyhalothrin (not lambda-cyhalothrin) to *Daphnia magna* were examined by Day (1991). Significantly less cyhalothrin was accumulated by *D. magna* when the DOC concentration was 3.1 mg/L or higher. The 48-hr EC₅₀ decreased with increasing DOC concentrations for all pyrethroids tested. For cyhalothrin, the 48-hr EC₅₀ increased 1.74-fold as the DOC increased from 1.3 mg/L DOC (EC₅₀=0.19 μ g/L) to 9.7 mg/L DOC (EC₅₀=0.33 μ g/L); the trends were more pronounced for other the pyrethroids deltamethrin and fenvalerate.

Smith and Lizotte (2007) conducted lambda-cyhalothrin and gamma-cyhalothrin toxicity tests with *Hyalella azteca* and twelve unfiltered pond waters with varying concentrations of the following four water quality parameters: turbidity, suspended solids (TSS), dissolved organic carbon (DOC), and chlorophyll a (chl a). The EC₅₀ values linearly increased as each parameter increased, indicating that bioavailability is directly related to the concentrations of these four parameters due to sorption of pyrethroids onto the particles, colloids or dissolved matter. The interaction of increased DOC and

phytoplankton (as chl a) decreased toxicity of lambda-cyhalothrin to H. azteca by more than 10-fold. The following equations were derived from linear regression of the concentration of the given parameter and the lambda-cyhalothrin EC_{50} values (ng/L):

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Turbidity (x, NTU): EC_{50} = 0.216x + 3.04, R^2 = 0.712, p = 0.0006 TSS (x, mg/L): EC_{50} = 0.179x + 3.15, R^2 = 0.644, p = 0.0017 DOC (x, mg/L): EC_{50} = 0.546x + 1.07, R^2 = 0.847, p < 0.0001 Chlorophyll a (x, µg/L): EC_{50} = 0.123x + 2.61, R^2 = 0.742, P = 0.0003
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These equations could be used to predict the EC_{50} for H. azteca when a particular water quality parameter is measured, but they cannot be used to determine compliance with the acute criterion because these relationships are not valid for multiple species. Yang $et\ al$. (2009) examined the effect of black carbon (BC) in the form of charcoal on lambdacyhalothrin accumulation in *Chironomus tentans*, and found that it had a very similar effect on bioavailability as sediment organic carbon. These studies suggest that the freely dissolved concentration will be the most accurate predictor of toxicity and that bound lambda-cyhalothrin was unavailable to the organisms that were studied.

As a counterpoint, equilibrium partitioning would suggest that as organisms take up lambda-cyhalothrin, more lambda-cyhalothrin will desorb from particles, so the fraction absorbed to solids is likely not completely unavailable. Benthic organisms, such a *Hyalella azteca*, may be at greater risk because of their exposure to porewater and close proximity to sediments. Additionally, the role of dietary exposure on bioavailability of pyrethroids has not been considered. Organisms living in contaminated waters are also ingesting food with sorbed hydrophobic compounds that can be desorbed by digestive juices (Mayer *et al.* 2001). The effects of dietary exposure may also be species-specific, depending on typical food sources; some species may have greater interaction with particles, increasing their exposure.

Section 3-5.1 of the methodology (TenBrook *et al.* 2009a) suggests that if studies indicate that fewer than three phases of the pesticide (sorbed to solids, sorbed to dissolved solids, or freely dissolved in the water) are bioavailable that compliance may be based on the concentration in the bioavailable phase(s). The studies above suggest that the freely dissolved fraction of lambda-cyhalothrin is the primary bioavailable portion of lambda-cyhalothrin, and that this concentration is the best indicator of toxicity. At this point, this recommendation is not being made for compliance assessment, but it is useful to consider how the freely dissolved concentration can be determined and how these methods compare to analytical methods used in toxicity test.

The most direct way to determine compliance would be to measure the lambda-cyhalothrin concentration in the dissolved phase to determine the total bioavailable concentration. Solid-phase microextraction (SPME) has shown to be the best predictor of pyrethroid toxicity in several studies (Bondarenko *et al.* 2007, Hunter *et al.* 2008, Xu *et al.* 2007, Yang 2006a, 2006b). Filtration of sediments is another option. Glass fiber filters with a nominal pore size of 0.7 μ m or 0.45 μ m are often used to remove the suspended sediments or both suspended sediments and dissolved organic matter, but the filters can

interfere with the detection of hydrophobic contaminants. Gomez-Gutierrez *et al.* (2007) found that adsorption to filters was positively correlated with the log K_{ow} and solubility values of the compounds, and that on average 58% of the one pyrethroid tested (a 50 ng/L solution of permethrin) was lost on the filter. This loss may be critical for determining compliance at environmental concentrations.

Alternately, the following equation can be used to translate total lambdacyhalothrin concentrations measured in water to the associated dissolved lambdacyhalothrin concentrations:

$$C_{dissolved} = \frac{C_{total}}{1 + ((K_{OC} \cdot [SS]) / foc) + (K_{DOC} \cdot [DOC])}$$
(1)

where: $C_{dissolved}$ = concentration of chemical in dissolved phase ($\mu g/L$);

 C_{total} = total concentration of chemical in water (μ g/L);

 K_{OC} = organic carbon-water partition coefficient (L/kg);

[SS] = concentration of suspended solids in water (kg/L);

 f_{oc} = fraction of organic carbon in suspended sediment in water;

[DOC] = concentration of dissolved organic carbon in water (kg/L);

 K_{DOC} = organic carbon-water partition coefficient (L/kg) for DOC.

To determine compliance by this calculation, a site specific K_{OC} and suspended sediment data are required, including the concentration and the fraction of organic carbon. The sorption of lambda-cyhalothrin to suspended solids and dissolved organic matter depend on the physical and chemical properties of the suspended solids resulting in a range of K_{OC} values (see section 3). This suggests that bioavailability may not be predicted based on a simple relationship and should not be estimated without site-specific data. Generating this site-specific data is fairly laborious, making SPME a more desirable choice.

While the literature suggests that the freely dissolved lambda-cyhalothrin concentrations are the most accurate predictor of toxicity, ten (of twenty) available toxicity values used to derive the acute criterion are based on nominal values, including the two lowest values for *Chaoborus* sp. and *Hyalella azteca*. These toxicity values are not measured whole-water concentrations or freely dissolved concentrations, by either of the methods described above. Several authors discuss that the use of nominal concentrations to calculate LC₅₀ values may considerably overestimate those values because there is likely considerable loss of pyrethroids to the sides of glass containers (Anderson *et al.* 2006, Day 1991, Froelich *et al.* 1984, Wheelock *et al.* 2005). Nominal toxicity values used in this report likely underestimate the sensitivity of organisms to lambda-cyhalothrin.

At this time we recommend that criteria compliance be based on whole-water lambda-cyhalothrin concentrations. Criteria based on nominal concentrations are likely to be underprotective and the role of dietary exposure has not been characterized; however, the use of whole-water concentrations is likely to be overprotective. The use of

whole-water lambda-cyhalothrin concentrations for compliance is currently the best way to ensure protection, compensating for the use of nominal concentrations and unknown effects of dietary exposure. This recommendation should be revised when more toxicity data based on measured concentrations are available.

10. Mixtures

Lambda-cyhalothrin often occurs in the environment with other pyrethroid pesticides (Werner & Moran 2008). Since compounds in this class have a similar mode of action, either the toxic unit or the relative potency factor approach can be used to determine compliance in cases where pyrethroid mixtures are present in environmental samples as presented in section 3-5.2.1 of the methodology (TenBrook *et al.* 2009a).

Piperonyl butoxide (PBO) is commonly added to pyrethroid insecticide treatments because it is known to increase the toxic effects of pyrethroids (Weston *et al.* 2006). No interaction coefficients (K) have been derived with relevant species to describe synergism between lambda-cyhalothrin and PBO. Consequently, it is not possible to quantify this non-additive toxicity and there is no accurate way to account for this interaction in compliance determination.

The effects on *Daphnia magna* mortality and feeding due to binary mixtures of lambda-cyhalothrin with deltamethrin, copper, and cadmium were examined in a study by Barata *et al.* (2006). The two concepts of concentration addition (CA) and independent action (IA) were used to predict mixture toxicity at various tested mixture ratios. Slight antagonism was observed in the lambda-cyhalothrin – deltamethrin mixture, which is unexpected because they have the same pharmacological mode of action. Neither method was able to consistently predict joint toxicity for the various mixtures. Another study by Barata *et al.* (2007) tested binary mixture toxicity of lambda-cyhalothrin and cadmium to *Daphnia magna* and examined reproductive effects. The joint toxicity of cadmium and lambda-cyhalothrin was equally predicted by the CA and IA models, even though these two chemicals do not have similar modes of pharmacological action, they do have similar ecotoxicological modes of action (Barata *et al.* 2007).

No studies on aquatic organisms were found in the literature that could provide a quantitative means to consider mixtures of lambda-cyhalothrin with other classes of pesticides. Although there are examples of non-additive toxicity for lambda-cyhalothrin and other chemicals, a multispecies interaction coefficient is not available for any chemical with lambda-cyhalothrin, and therefore the concentrations of non-additive chemicals cannot be used for criteria compliance (section 3-5.2.2, TenBrook *et al.* 2009a).

11. Temperature, pH, other water quality effects

Temperature has been found to be inversely proportional to the aquatic toxicity and bioavailability of pyrethroids (Miller & Salgado 1985, Werner & Moran 2008). In

fact, the increase of toxicity of pyrethroids with decreasing temperature has been used to implicate pyrethroids as the source of toxicity in environmental samples (Phillips *et al.* 2004). The inverse relationship between temperature and pyrethroid toxicity is likely due to the increased sensitivity of an organism's sodium channels at low temperatures (Narahashi *et al.* 1998).

The toxicity of sediments contaminated with pyrethroids (including lambdacyhalothrin) was more than twice as toxic when tested at 18 °C compared to 23 °C (Weston *et al.* 2008). The enhanced toxic effects of pyrethroids at lower temperatures may not be as accurately represented by the results of typical laboratory toxicity tests, which tend to be run at warmer temperatures, 20-23 °C (USEPA 1996a, USEPA 1996b, USEPA 2000), than those of the habitats of coldwater fishes, about 15 °C or lower (Sullivan *et al.* 2000).

In studies that used topical exposures (more relevant to spray application exposure to target a pest), the difference in toxicity can increase by a factor of about 1.5 to a factor of 10, in the temperature range of about 10 to 27 °C (Kumaraguru & Beamish 1981; Punzo 1993; Schnitzerling 1985). A simple relationship of temperature and the binding of pyrethroids to a site of action may account for the increase of toxicity for permethrin to the cattle tick *Boophilus-microplus* (Schnitzerling 1985).

Unfortunately, there is limited data in this regard using aquatic exposures with aquatic species, making it infeasible to quantify the relationship between the toxicity of lambda-cyhalothrin and temperature for water quality criteria at this time (section 3-5.3, TenBrook *et al.* 2009a). Several studies that examined the effects of DOC, suspended solids, turbidity, and chlorophyll a concentrations are discussed in the bioavailability section 9 above. No other studies on lambda-cyhalothrin were found that examined the effects of pH or other water quality parameters on toxicity, thus, there is no way to incorporate any of these parameters into criteria compliance.

12. Sensitive species

The derived acute criterion (1 ng/L) is below all of the acute values in the available data sets. The lowest acute value in the data sets rated RR, RL, LR, or LL (Tables 3 - 5) is 2.3 ng/L for the amphipod *Hyalella azteca* (Hamer *et al.* 1998). The derived chronic criterion (1 ng/L) is protective of the lowest chronic value in the data set rated RR (Tables 6 and 7), which is an MATC of 2.63 ng/L for *Daphnia magna* (Farrelly & Hamer 1989). The lowest chronic value in the data set rated RL, LR, or LL (Table 9) is an MATC of 0.32 ng/L for the mysid shrimp *Mysidopsis bahia*, a saltwater species. As the goal of this method is to protect freshwater species, and saltwater species may have different sensitivities to lambda-cyhalothrin than freshwater species, it is therefore not recommended to adjust the chronic criterion downward to be protective of this species.

13. Ecosystem and other studies

Eleven mesocosm, microcosm or ecosystem (field and laboratory) studies were identified and rated (Table 10). Five of the studies were rated relevant and reliable (R; Farmer *et al.* 1995, Roessink *et al.* 2005, Schroer *et al.* 2004, Van Wijngaarden *et al.* 2006, Wendt-Rasch *et al.* 2004), and three were rated less relevant and reliable (L; Gu *et al.* 2007, Lauridsen & Friberg 2005, Rasmussen *et al.* 2008) and are used as supporting data. These studies were primarily outdoor microcosms mimicking small riverine environments and all exposures used commercial formulations of lambda-cyhalothrin. Several studies report a community NOEC to which the calculated criteria may be compared.

Several studies reported significant macroinvertebrate mortality and drift due to exposure to lambda-cyhalothrin (Farmer *et al.* 1995, Lauridsen and Friberg 2005, Rasmussen *et al.* 2008, Wendt-Rasch *et al.* 2004). Gammarus species were examined in several studies and it was found that they were particularly sensitive to lambda-cyhalothrin. Phytoplankton and algae productivity increased in response to lambda-cyhalothrin exposure (Farmer *et al.* 1995 Rasmussen *et al.* 2008, Wendt-Rasch *et al.* 2004). This decrease in macroinvertebrate populations was the likely cause of the increase in phytoplankton and algae, as these organisms are known to graze on algae.

Roessink *et al.* 2005, Schroer *et al.* 2004, and Van Wijngaarden *et al.* 2006 examined the effects of lambda-cyhalothrin on macroinvertebrates in ditch microcosm systems in two seasons (spring and late summer) with two types of vegetation (eutrophic and mesotrophic), and compared them to laboratory tests for the same species. Van Wijngaarden *et al.* (2006) and Roessink *et al.* (2005) report various community-level NOEC values depending on the season and trophic system, the lowest being < 10 μg/L, and Schroer *et al.* (2004) reports a community-level NOEC of 10 ng/L. Schroer *et al.* (2004) also calculated a community-level criterion of 4.1 ng/L, while the criterion calculated based on laboratory single-species data was 2.7 ng/L. The derived acute and chronic criteria are below all of the reported NOEC values for this set of studies. The lowest community-level NOEC reported was 10 ng/L, which is a factor of ten larger than the derived criteria.

Lauridsen and Friberg 2005 (rated L) examined macroinvertebrate drift in outdoor experimental channels with two insects (*Baetis rhodani* and *Leuctra fusca/digitata*) and the amphipod *Gammarus pulex*. Catastrophic drift was observed for all three species during the one hour pulse exposure and 2-3 h post-exposure. *G. pulex* was significantly affected at 0.001 µg/L (nominal), which is equivalent to the derived criteria, and the insects were affected at 0.01 µg/L. It is not recommended to adjust the criteria downward to be protective of possible adverse affects indicated by this study because an acute toxicity value for *G. pulex* was included in the criteria derivation, and the reported concentration is nominal, and the measured concentration may have been significantly lower.

14. Threatened and endangered species

Current lists of state and federally listed threatened and endangered plant and animal species in California were obtained from the California Department of Fish and Game website (http://www.dfg.ca.gov/biogeodata/cnddb/pdfs/TEAnimals.pdf; CDFG 2008). Two listed animal species are represented in the dataset. Five Evolutionarily Significant Units of *Oncorhynchus mykiss* are listed as federally threatened or endangered throughout California. The acute data set includes a SMAV for *O. mykiss* of 0.27 µg/L calculated from three studies rated RR. The unarmored threespine stickleback (*Gasterosteus aculeatus williamsoni*) is represented in the RR data set with a with an LC₅₀ value of 0.40 (0.33-0.50) µg/L reported for *G. aculeatus*. Both of these values in the data set were included in the acute criterion calculation and are well above the recommended acute criterion.

The USEPA interspecies correlation estimation (ICE v. 1.0; Raimondo *et al.* 2007) software was used to estimate toxicity values for the listed animals or plants represented in the acute data set by members of the same family or genus. Table 11 summarizes the results of the ICE analyses. The values in Table 11 range from 0.277 μ g/L for Coho salmon to 0.539 μ g/L for Chinook salmon.

No plant studies used in the criteria derivation were of state or federal endangered, threatened or rare species. There are no aquatic plants listed as state or federal endangered, threatened or rare species so they are not considered in this section.

Based on the available data and estimated values for animals, there is no evidence that the calculated acute and chronic criteria will be under-protective of threatened and endangered species.

15. Bioaccumulation

Lambda-cyhalothrin has a log K_{ow} of 7.0 and a molecular weight of 449.85 (section 3), which indicates its bioaccumulative potential (section 3-7.1, TenBrook *et al.* 2009a). No biomagnification factor (BMF) values were found in the literature for lambda-cyhalothrin. Bioaccumulation of lambda-cyhalothrin has been measured in several studies (Table 1), which are briefly summarized here. The bioconcentration factor (BCF) in carp (*Cyprinus carpio*) varied from 850-7340 depending on what portion of the fish was analyzed (Yamauchi *et al.* 1984). The BCF values for channel catfish (*Ictalurus punctatus*) were lower than those for carp, ranging 7-66 depending on which portion was analyzed (Hamer and Hill 1985). Bioconcentration was examined by Hamer *et al.* (1999) in *Chironomus riparius* in water only and water-sediment systems and the BCF values were very similar for the two systems (2000 and 2300, respectively). The BCF for *Daphnia magna* was significantly lower than those for *C. riparius* at 194 (Hamer and Hill 1985).

To check that these criteria are protective of terrestrial wildlife that may consume aquatic organisms, a bioaccumulation factor (BAF) was used to estimate the water concentration that would roughly equate to a reported toxicity value for consumption of fish by terrestrial wildlife. These calculations are further explained in section 3-7.1 of the

methodology (TenBrook *et al.* 2009a). The BAF of a given chemical is the product of the bioconcentration factor (BCF) and a biomagnification factor (BMF), such that BAF=BCF*BMF. For a conservative estimate, the BCF value of 2240 L/kg for *Cyprinus carpio* was used (Table 1). A default BMF value of 10 was chosen based on the log K_{ow} of lambda-cyhalothrin (Table 3.15, TenBrook *et al.* 2009a). An oral predator dietary LC₅₀ value for mallard duck of 3948 mg/kg feed (Roberts *et al.* 1985) was used in the calculation. The NOEL value reported by Beavers *et al.* (1990) was not used because it is likely an underestimation because there were no effects observed at any of the tested concentrations.

$$NOEC_{water} = \frac{LC_{50,oral_predator}}{BCF_{food_item} * BMF_{food_item}}$$

Mallard:
$$NOEC_{water} = \frac{3948 \frac{mg}{kg}}{2240 \frac{l}{kg} * 10} = 0.176 \frac{mg}{L} = 176 \frac{\mu g}{L}$$

In this example, the calculated chronic criterion is 176-fold below the estimated NOEC_{water} value for the mallard and adverse effects due to bioaccumulation. are not expected.

To check that these criteria are protective of humans that may consume aquatic organisms, a BAF will be used to estimate the water concentration that would roughly equate to a limit for human food consumption. An appropriate BAF was not available in the data set. The BCF value for carp muscle of 850 (Yamauchi *et al.* 1985, Table 1) and a human food tolerance level are used. There are no tolerance or FDA action levels for fish tissue (USFDA 2000), but there is a food tolerance for hog meat at 0.1 ppm (USEPA 2007). This value can be used to roughly estimate if bioconcentration could cause lambda-cyhalothrin concentrations in fish tissues to be of concern to human heath.

Human:
$$NOEC_{water} = \frac{0.1^{mg/kg}}{850 \frac{L}{kg} * 10} = 0.0000176^{mg/L} = 0.0176^{\mu g/L} = 18^{ng/L}$$

In this example, the derived chronic criterion of 1 ng/L is more than an order of magnitude below the estimated water concentrations of concern for wildlife and humans (176,000 ng/L and 18 ng/L). Therefore, adhering to the derived lambda-cyhalothrin criteria should not conflict with other efforts to protect wildlife or human health from lambda-cyhalothrin exposure.

16. Harmonization/coherence across media

This section addresses how the maximum allowable concentration of lambdacyhalothrin might impact life in other environmental compartments through partitioning (section 3-7.2, TenBrook *et al.* 2009a). However, there are no federal or state sediment or air quality standards for lambda-cyhalothrin (California Air Resources Board 2005, California Department of Water Resources 1995, USEPA 2006a, USEPA 2006b,) to enable this kind of extrapolation. For biota, the limited data on bioconcentration or biomagnification of lambda-cyhalothrin was addressed in the bioaccumulation section (section 15).

17. Assumptions, Limitations and Uncertainties

The assumptions, limitations and uncertainties involved in criteria derivation should be available to inform environmental managers of the accuracy and confidence in the derived criteria (section 3-8.0, TenBrook *et al.* 2009a). Chapter 2 of the methodology discusses these points for each section as different procedures were chosen, such as the list of assumptions associated with using a species sensitivity distribution (SSD), included in section 2-3.1.5.1, and reviews the assumptions in section 2-7.0 (TenBrook *et al.* 2009a). The different calculations of distributional estimates included in section 7 of this report may be used to consider the uncertainty in the resulting acute criterion.

For lambda-cyhalothrin, the major limitation was in the chronic toxicity data set. Three of five taxa requirements were not met (the salmonid, benthic crustacean and insect), which precluded the use of a SSD; therefore, an acute to chronic ratio (ACR) was used to derive the chronic criterion. There was measured data available for calculation of a multi-species ACR (as specified in section 3-4.2.1, TenBrook *et al.* 2009a). Particularly of concern for the chronic toxicity data set was the lack of data on *Hyalella azteca*, which was the most sensitive species in the acute toxicity data set.

Another concern that could not be accounted for quantitatively with the acute and chronic criteria is the increase in toxicity from lower temperatures. Most of the toxicity data were from tests performed at standard temperature, usually around 20 °C. However, many streams in the California Central Valley often have lower water temperatures. If colder water bodies are impacted by concentrations of lambda-cyhalothrin, it may be appropriate to apply an additional safety factor to the lambda-cyhalothrin criteria for those areas, to ensure adequate protection. A rough factor of two could be estimated from a study by Weston *et al.* (2008), however, a study relating temperature to toxicity of lambda-cyhalothrin in *Hyalella azteca* would be ideal to derive such an adjustment factor.

18. Comparison to National Standard Methods

This section is provided as a comparison between the new methodology for criteria calculation (TenBrook *et al.* 2009a) and the current USEPA (1985) national standard. The following example lambda-cyhalothrin criteria were generated using the USEPA 1985 methodology with the data set generated in this lambda-cyhalothrin criteria report.

The USEPA acute methods have three additional taxa requirement beyond the five required by the methodology used in this criteria report (section 3-3.1, TenBrook *et al.* 2009a). They are:

- 1. A third family in the phylum Chordata (e.g., fish, amphibian);
- 2. A family in a phylum other than Arthropoda or Chordata (e.g., Rotifera, Annelida, Mollusca);
- 3. A family in any order of insect or any phylum not already represented.

Two out of the three of these additional requirements are met as follows:

- 1. The other fish/amphibian requirement is met with data from zebra danio or any of six other fish species available.
- 2. This requirement not met because all data are from organisms in the phylum Arthropoda or Chordata.
- 3. This requirement is met because *Cloeon dipterum* is an insect in a different family than *Chaoborus* sp.

Strictly speaking, the USEPA methodology cannot be used to calculate an acute criterion for lambda-cyhalothrin. However, since the California Department of Fish and Game have used data sets that met only seven of eight requirements in the USEPA methodology, this will be done here.

Using the log-triangular calculation (following the USEPA 1985 guidelines) and the lambda-cyhalothrin data set from Table 3 containing twenty species values, the following criterion was calculated (Note: USEPA methodology uses *genus* mean acute values, while *species* mean acute values are used in this methodology and are reported in Table 3. Since there is only one species from each genus in Table 3, this final data set would be the same in both schemes.):

```
Example Final Acute Value (5<sup>th</sup> percentile value) = 0.001845 \mu g/L
```

```
Example Acute Criterion = final acute value \div 2 = 0.001845 \mug/L \div 2 = 0.000922 \mug/L = 0.9 ng/L
```

For the chronic criterion, the lambda-cyhalothrin data set only has data from 2 species, which are not enough for use in a species sensitivity distribution by either method. The USEPA 1985 methodology contains a similar acute to chronic ratio (ACR) procedure as in the methodology used in this criteria report, to be used when three acceptable ACRs are available. The same three ACR values calculated for this methodology (Table 8) were calculated according to the USEPA 1985 methodology to give a final ACR of 4.73. The chronic criterion is calculated by dividing the Final Acute Value (FAV) by the Final ACR:

Example Chronic Criterion = final acute value ÷ final ACR

=
$$0.001845 \ \mu g/L \div 4.73 = 0.00039 \ \mu g/L$$

= $0.4 \ ng/L$

19. Final criteria statement

The final criteria statement is:

Aquatic life in the Sacramento River and San Joaquin River basins should not be affected unacceptably if the four-day average concentration of lambda-cyhalothrin does not exceed 0.001 μ g/L (1 ng/L) more than once every three years on the average and if the one-hour average concentration does not exceed 0.001 μ g/L (1 ng/L) more than once every three years on the average.

To date, there are no established criteria for lambda-cyhalothrin to which the criteria calculated in this report can be compared except those example criteria calculated by the USEPA 1985 method in the above section. The example acute criterion calculated by the USEPA 1985 method is almost identical to the acute criterion derived using this new methodology, and the example chronic criterion calculated by the USEPA 1985 method is lower than the chronic criterion derived using the new methodology by approximately a factor of 2. Solomon *et al.* (2001) performed a probabilistic risk assessment with pyrethroids. Saltwater and freshwater toxicity data were combined so the lowest toxicity value in the data set was 4 ng/L (for mysid, a saltwater species). The 5th percentile value for lambda-cyhalothrin, based on a log-normal distribution, was < 4 ng/L, although much of the author's discussion centered on the 10th percentile as the protective limit, which was 10 ng/L for lambda-cyhalothrin. For compounds that had larger toxicity data sets, separate analyses were performed for freshwater and saltwater data. Differences were found especially for invertebrates, which suggested that the risk to freshwater and saltwater organisms should be assessed separately.

The derived criteria appear to be protective considering bioaccumulation, ecosystem level toxicity and threatened and endangered species as discussed above in the report, but the criteria calculations should be updated whenever new data is available.

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Data Tables

Table 3. Final acute toxicity data set for lambda-cyhalothrin. All studies were rated RR and were conducted at standard temperature. S: static; SR: static renewal; FT: flow-through.

Species	Common Identifier	Family	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (μg/L) (95% CI)	Reference
Asellus aquaticus	Isopod	Aselloidea	S	Nom	88.0%	48 h	20	Immobility	NR	0.026 (0.018-0.036)	Hamer et al. 1998
Brachydanio rerio	Zebra danio	Cyprinidae	FT	Meas	88.7%	96 h	25	Mortality	0.70 g, 36 mm	0.64 (0.48-0.90)	Kent & Shillabeer 1997c
Ceriodaphnia dubia	Daphnid	Daphniidae	S	Nom	97.0%	48 h	25	Mortality	< 24 h	0.200 ± 0.090	Wheelock et al. 2004
Chaoborus sp.	Phantom midge	Chaoboridae (Insecta)	S	Nom	88.0%	48 h	20	Maintenance of body shape/equil.	larvae	0.0028 (0.0018-0.0041)	Hamer et al. 1998
Cloeon dipterum	Mayfly nymph	Baetidae (Insecta)	S	Nom	88.0%	48 h	20	Immobility	nymph	0.038 (0.023-0.093)	Hamer et al. 1998
Corixa sp.	Hemipteran	Corixidae	S	Nom	88.0%	48 h	20	Immobility	NR	0.030 (0.021-0.042)	Hamer et al. 1998
Cyclops sp.	Copepod	Cyclopidae	S	Nom	88.0%	48 h	20	Immobility	NR	0.300 (0.200-0.460)	Hamer et al. 1998
Daphnia magna	Daphnid	Daphniidae	FT	Meas	94.3%	72 h	20	Mortality	< 24 h	0.013 (0.010-0.017)	Farrelly & Hamer 1989
Gammarus pulex	Amphipod	Gammaridae	FT	Meas	99.2%	96 h	15	Immobility	5 mm, > 3 weeks old	0.0059	Hamer et al. 1985a
Gasterosteus aculeatus	3 spined stickleback	Gasterosteidae	FT	Meas	87.7%	96 h	12	Mortality	0.41 g, 34 mm	0.40 (0.33-0.50)	Long & Shillabeer 1997a
Hyalella azteca	Amphipod	Hyalellidae	S	Nom	88.0%	48 h	20	Immobility	NR	0.0023 (0.0010-0.0078)	Hamer et al. 1998

Species	Common Identifier	Family	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (μg/L) (95% CI)	Reference
Hydracarina (Class)	Water mite	NR	S	Nom	88.0%	48 h	20	Immobility	NR	0.047 (0.033-0.062)	Hamer et al. 1998
Ictalurus punctatus	Channel catfish	Ictaluridae	FT	Meas	87.7%	96 h	17	Mortality	1.57 g, 48 mm	0.16 (0.13-0.20)	Long & Shillabeer 1997b
Lepomis macrochirus Rafinesque	Bluegill sunfish	Centrarchidae	FT	Meas	99.0%	96 h	21.9	Mortality	juvenile	0.106 (0.0855-0.140)	Marino & Rick 2001
Lepomis macrochirus	Bluegill sunfish	Centrarchidae	FT	Meas	98.0%	96 h	22	Mortality	1.51 g, 38.2 mm	0.21 (0.18-0.25)	Hill 1984b
Geomean										0.15	
Leuciscus idus	Golden orfe	Cyprinidae	FT	Meas	88.7%	96 h	12	Mortality	2.15 g, 53 mm	0.078 (0.056-0.11)	Kent & Shillabeer 1997a
Oncorhynchus mykiss	Rainbow trout	Salmonidae	FT	Meas	99.0%	96 h	12	Mortality	39 mm, 0.52 g	0.19 (0.16-0.20)	Machado 2001b
Oncorhynchus mykiss	Rainbow trout	Salmonidae	FT	Meas	81.5%	96 h	12	Mortality	43 mm, 1.12 g	0.44 (0.38-0.51)	Tapp et al. 1989
Oncorhynchus mykiss	Rainbow trout	Salmonidae	FT	Meas	98.0%	96 h	12	Mortality	38.3mm, 0.83 g	0.24 (0.08-0.70)	Hill 1984a
Geomean										0.27	
Ostracoda (class)	Seed shrimp	NR	S	Nom	88.0%	48 h	20	Immobility	NR	3.300 (2.100-6.600)	Hamer et al. 1998
Pimephales promelas	Fathead minnow	Cyprinidae	FT	Meas	97.0%	96 h	25	Mortality	larvae	0.360 (0.252-0.765)	Tapp et al. 1990
Pimephales promelas	Fathead minnow	Cyprinidae	FT	Meas	88.7%	96 h	25	Mortality	0.37 g, 28 mm	0.70 (0.38-1.3)	Kent & Shillabeer 1997e
Geomean										0.50	
Poecilia reticulata	Guppy	Poeciliidae	FT	Meas	88.7%	96 h	25	Mortality	0.62 g, 33 mm	2.3 (1.8-3.1)	Kent & Shillabeer 1997b

	Species	Common Identifier	Family	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (μg/L) (95% CI)	Reference
1	Procambarus clarkii	Crayfish	Cambaridae	SR	Nom	99.1%	96 h	21.7	Mortality	3 months old	0.16 (0.06-0.27)	Barbee & Stout 2009

Table 4. Reduced acute data rated RR with given reason for exclusion. S: static; SR: static renewal; FT: flow-through.

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (µg/L) (95% CI)	Reference	Reason
Brachydanio rerio	Zebra danio	FT	Meas	88.7%	24 h	25	Mortality	0.70 g, 36 mm	0.97 (0.74-1.4)	Kent & Shillabeer 1997c	A
Brachydanio rerio	Zebra danio	FT	Meas	88.7%	48 h	25	Mortality	0.70 g, 36 mm	0.80 (0.62-1.1)	Kent & Shillabeer 1997c	A
Brachydanio rerio	Zebra danio	FT	Meas	88.7%	72 h	25	Mortality	0.70 g, 36 mm	0.64 (0.48-0.90)	Kent & Shillabeer 1997c	A
Daphnia magna	Daphnid	S	Meas	99.0%	48 h	NR	Immobility	4th instar juveniles	0.39 (0.38-0.40)	Barata et al. 2006	В
Daphnia magna	Daphnid	S	Meas	96.5%	24 h	20	Immobility	< 24 h	5.04	Farrelly et al.1984	В
Daphnia magna	Daphnid	S	Meas	96.5%	48 h	20	Immobility	< 24 h	0.36	Farrelly et al.1984	В
Daphnia magna	Daphnid	SR	Meas	99.0%	48 h	21	Immobility	≤ 24 h	0.051 (0.034-0.10)	Machado 2001a	A
Gammarus pulex	Amphipod	S	Nom	88.0%	48 h	20	Immobility	NR	0.014 (0.0091-0.019)	Hamer et al. 1998	В
Gammarus pulex	Amphipod	FT	Meas	99.2%	24 h	15	Immobility	5 mm, > 3 weeks old	0.0102	Hamer et al. 1985a	A
Gammarus pulex	Amphipod	FT	Meas	99.2%	48 h	15	Immobility	5 mm, > 3 weeks old	0.008	Hamer et al. 1985a	A
Gammarus pulex	Amphipod	FT	Meas	99.2%	72 h	15	Immobility	5 mm, > 3 weeks old	0.0064	Hamer et al. 1985a	A
Gammarus pulex	Amphipod	FT	Meas	99.2%	24 h	15	Mortality	5 mm, > 3 weeks old	0.665	Hamer et al. 1985a	C
Gammarus pulex	Amphipod	FT	Meas	99.2%	48 h	15	Mortality	5 mm, > 3 weeks old	0.0712	Hamer et al. 1985a	C
Gammarus pulex	Amphipod	FT	Meas	99.2%	72 h	15	Mortality	5 mm, > 3 weeks old	0.0313	Hamer et al. 1985a	C
Gammarus pulex	Amphipod	FT	Meas	99.2%	96 h	15	Mortality	5 mm, > 3 weeks old	0.0127	Hamer et al. 1985a	С
Gasterosteus aculeatus	3 spined stickleback	FT	Meas	87.7%	24 h	12	Mortality	0.41 g, 34 mm	0.73 (0.68-0.79)	Long & Shillabeer 1997a	A

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (µg/L) (95% CI)	Reference	Reason
Gasterosteus aculeatus	3 spined stickleback	FT	Meas	87.7%	48 h	12	Mortality	0.41 g, 34 mm	0.44 (0.36-0.56)	Long & Shillabeer 1997a	A
Gasterosteus aculeatus	3 spined stickleback	FT	Meas	87.7%	72 h	12	Mortality	0.41 g, 34 mm	0.43 (0.35-0.54)	Long & Shillabeer 1997a	A
Ictalurus punctatus	channel catfish	FT	Meas	87.7%	24 h	17	Mortality	1.57 g, 48 mm	0.82 (0.67-11)	Long & Shillabeer 1997b	A
Ictalurus punctatus	channel catfish	FT	Meas	87.7%	48 h	17	Mortality	1.57 g, 48 mm	0.43 (0.25-0.73)	Long & Shillabeer 1997b	A
Ictalurus punctatus	channel catfish	FT	Meas	87.7%	72 h	17	Mortality	1.57 g, 48 mm	0.18 (0.15-0.23)	Long & Shillabeer 1997b	A
Lepomis macrochirus Rafinesque	Bluegill sunfish	FT	Meas	99.0%	24 h	21.9	Mortality	juvenile	0.224 (0.152-1.742)	Marino & Rick 2001	A
Lepomis macrochirus Rafinesque	Bluegill sunfish	FT	Meas	99.0%	48 h	21.9	Mortality	juvenile	0.124 (0.0944-0.163)	Marino & Rick 2001	A
Lepomis macrochirus Rafinesque	Bluegill sunfish	FT	Meas	99.0%	72 h	21.9	Mortality	juvenile	0.118 (0.0944- 0.155)	Marino & Rick 2001	A
Lepomis macrochirus	Bluegill sunfish	FT	Meas	98.0%	24 h	22	Mortality	1.51 g, 38.2 mm	0.45 (0.38-0.52)	Hill 1984b	A
Lepomis macrochirus	Bluegill sunfish	FT	Meas	98.0%	48 h	22	Mortality	1.51 g, 38.2 mm	0.28 (0.23-0.32)	Hill 1984b	A
Lepomis macrochirus	Bluegill sunfish	FT	Meas	98.0%	72 h	22	Mortality	1.51 g, 38.2 mm	0.28 (0.23-0.32)	Hill 1984b	A
Leuciscus idus	golden orfe	FT	Meas	88.7%	72 h	12	Mortality	2.15 g, 53 mm	0.078 (0.056-0.11)	Kent & Shillabeer 1997a	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	99.0%	48 h	12	Mortality	39 mm, 0.52 g	0.29 (0.25-0.33)	Machado 2001b	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	99.0%	72 h	12	Mortality	39 mm, 0.52 g	0.22 (0.20-0.38)	Machado 2001b	A

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (µg/L) (95% CI)	Reference	Reason
Oncorhynchus mykiss	Rainbow trout	FT	Meas	81.5%	48 h	12	Mortality	43 mm, 1.12 g	0.57 (0.50-0.66)	Tapp et al. 1989	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	81.5%	72 h	12	Mortality	43 mm, 1.12 g	0.49 (0.43-0.58)	Tapp et al. 1989	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	98.0%	24 h	12	Mortality	38.3mm, 0.83 g	0.52 (0.46-0.60)	Hill 1984a	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	98.0%	48 h	12	Mortality	38.3mm, 0.83 g	0.40 (0.35-0.45)	Hill 1984a	A
Oncorhynchus mykiss	Rainbow trout	FT	Meas	98.0%	72 h	12	Mortality	38.3mm, 0.83 g	0.27 (0.09-0.80)	Hill 1984a	A
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	72 h	25	Mortality	larvae	0.407 (0.316-0.675)	Tapp et al. 1990	A
Pimephales promelas	Fathead minnow	FT	Meas	88.7%	24 h	25	Mortality	0.37 g, 28 mm	0.89 (0.73-1.1)	Kent & Shillabeer 1997e	A
Pimephales promelas	Fathead minnow	FT	Meas	88.7%	48 h	25	Mortality	0.37 g, 28 mm	0.89 (0.73-1.1)	Kent & Shillabeer 1997e	A
Pimephales promelas	Fathead minnow	FT	Meas	88.7%	72 h	25	Mortality	0.37 g, 28 mm	0.70 (0.38-1.3)	Kent & Shillabeer 1997e	A
Poecilia reticulata	guppy	FT	Meas	88.7%	24 h	25	Mortality	0.62 g, 33 mm	2.9 (1.6-5.1)	Kent & Shillabeer 1997b	A
Poecilia reticulata	guppy	FT	Meas	88.7%	48 h	25	Mortality	0.62 g, 33 mm	2.9 (1.6-5.1)	Kent & Shillabeer 1997b	A
Poecilia reticulata	guppy	FT	Meas	88.7%	72 h	25	Mortality	0.62 g, 33 mm	2.5 (1.9-3.4)	Kent & Shillabeer 1997b	A

Reduction Reasons

A. Not the most sensitive or appropriate duration

B. FT test preferred over S

C. Not the most sensitive endpoint

Table 5. Excluded acute data rated RL, LR, LL with rating and reason for exclusion given below. S: static; SR: static renewal; FT: flow-through. NR: not reported.

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (μg/L) (95% CI)	Reference	Rating/ Reason
Brachydanio rerio	Zebra fish	SR	Nom	formulation	24 h	25	Mortality	30-45 d old	8.26 (5.93-11.51)	Wang et al. 2007	LL 1, 7
Brachydanio rerio	Zebra fish	SR	Nom	formulation	48 h	25	Mortality	30-45 d old	3.91 (2.62-5.84)	Wang et al. 2007	LL 1, 7
Brachydanio rerio	Zebra fish	SR	Nom	formulation	72 h	25	Mortality	30-45 d old	2.05 (1.40-3.01)	Wang et al. 2007	LL 1, 7
Brachydanio rerio	Zebra fish	SR	Nom	formulation	96 h	25	Mortality	30-45 d old	1.94 (1.33-2.84)	Wang et al. 2007	LL 1, 7
Caridina laevis	Atyid shrimp	S	Nom	formulation	24 h	26	Mortality	Adult, 15- 20 mm	0.87 (0.76-0.98)	Sucahyo et al. 2008	RL 1, 7
Caridina laevis	Atyid shrimp	S	Nom	formulation	96 h	26	Mortality	Adult, 15- 20 mm	0.33 (0.30-0.37)	Sucahyo et al. 2008	RL 1, 7
Channa punctatus	Snakehead fish	SR	Nom	5.0%	96 h	27	Mortality	Teleost, 11-3 cm, 23 g	7.92	Kumar et al. 2007	LL 1, 7
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.5%	24 h	22	Mortality	0.60 g, 27.4 mm	1.34	Hill 1985	RL 2
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.5%	48 h	22	Mortality	0.60 g, 27.4 mm	1.14	Hill 1985	RL 2
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.5%	72 h	22	Mortality	0.60 g, 27.4 mm	0.85	Hill 1985	RL 2
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.5%	96 h	22	Mortality	0.60 g, 27.4 mm	0.81	Hill 1985	RL 2
Ischnura elegans	Damselfly nymph	S	Nom	88.0%	48 h	20	Immobility	nymph	0.130 (0.092-0.190)	Hamer et al. 1998	RL 1, 4
Macrobrachium nippoensis	shrimp	SR	Nom	formulation	24 h	16	Mortality	90 d old, 5.0 g, 4.5 cm	0.05 (0.04-0.07)	Wang et al. 2007	LL 1, 7

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (µg/L) (95% CI)	Reference	Rating/ Reason
Macrobrachium nippoensis	shrimp	SR	Nom	formulation	48 h	16	Mortality	90 d old, 5.0 g, 4.5 cm	0.05 (0.04-0.06)	Wang et al. 2007	LL 1, 7
Macrobrachium nippoensis	shrimp	SR	Nom	formulation	72 h	16	Mortality	90 d old, 5.0 g, 4.5 cm	0.04 (0.03-0.06)	Wang et al. 2007	LL 1, 7
Macrobrachium nippoensis	shrimp	SR	Nom	formulation	96 h	16	Mortality	90 d old, 5.0 g, 4.5 cm	0.04 (0.03-0.05)	Wang et al. 2007	LL 1, 7
Mysidopsis bahia	mysid shrimp	FT	Meas	97.0%	24 h	25	Mortality	<48 h	> 0.017	Thompson 1985	RL 2, 4
Mysidopsis bahia	mysid shrimp	FT	Meas	97.0%	48 h	25	Mortality	<48 h	0.0075 (0.0061- 0.0096)	Thompson 1985	RL 2, 4
Mysidopsis bahia	mysid shrimp	FT	Meas	97.0%	72 h	25	Mortality	<48 h	0.0049 (0.0041- 0.0058)	Thompson 1985	RL 2, 4
Mysidopsis bahia	mysid shrimp	FT	Meas	97.0%	96 h	25	Mortality	<48 h	0.0041 (0.0034- 0.0049)	Thompson 1985b+L7	RL 2, 4
Oryzias latipes	Japanese rice fish	FT	Meas	88.7%	24 h	25	Mortality	0.22g, 25 mm	2.1 (1.5-3.3)	Kent & Shillabeer 1997d	LR 3
Oryzias latipes	Japanese rice fish	FT	Meas	88.7%	48 h	25	Mortality	0.22g, 25 mm	1.5 (1.0-2.6)	Kent & Shillabeer 1997d	LR 3
Oryzias latipes	Japanese rice fish	FT	Meas	88.7%	72 h	25	Mortality	0.22g, 25 mm	1.4 (0.93-2.3)	Kent & Shillabeer 1997d	LR 3

Species	Common Identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/size	LC/EC ₅₀ (μg/L) (95% CI)	Reference	Rating/ Reason
Oryzias latipes	Japanese rice fish	FT	Meas	88.7%	96 h	25	Mortality	0.22g, 25 mm	1.4 (0.93-2.3)	Kent & Shillabeer 1997d	LR 3

Exclusion Reasons

- 1. Not a standard method
- 2. Saltwater
- 3. Family not found in N. America
- 4. Unacceptable control response
- 5. Control response not reported
- 6. Low reliability score
- 7. Low chemical purity

95% CI: 95% confidence interval

Table 6. Final chronic animal toxicity data set for lambda-cyahlothrin. All studies were rated RR. S: static; SR: static renewal; FT: flow-through. NR: not reported

Species	Common identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/ size	NOEC (µg/L)	LOEC (µg/L)	MATC (μg/L)	Reference
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Reproduction (young/female/d)	< 24 h	0.00198	0.00350	0.00263	Farrelly & Hamer 1989
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Reproduction (young/female/d)	< 24 h	0.00375	0.00490	0.00429	Hamer et al. 1985b
											0.00336	
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	56 d	25	F1 Survival	F1 larvae	0.031	0.062	0.044	Tapp et al. 1990

Table 7. Acceptable reduced chronic data rated RR with reason for exclusion given below. S: static; SR: static renewal; FT: flow-through. NR: not reported

unot	ign. NK: i	ioi iep	orieu										
Species	Common name	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/ size	NOEC (µg/L)	LOEC (µg/L)	MATC (μg/L)	Reference	Reason
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Reproduction (total young)	<24 h	0.00198	0.00350	0.00263	Farrelly & Hamer 1989	
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Reproduction (total young)	<24 h	0.00490	0.00850	0.00645	Hamer et al. 1985b	
											0.00412		A
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Reproduction (# of female repro days)	<24 h	0.00850	0.01830	0.01247	Hamer et al. 1985b	A
Daphnia magna	Daphnid	FT	Meas	94.3%	9 d	20	Length	<24 h	0.01830	0.03720	0.02609	Hamer et al. 1985b	
Daphnia magna	Daphnid	FT	Meas	94.3%	21 d	20	Length	<24 h	0.00937	0.01910	0.01338	Farrelly & Hamer 1989	
											0.0187		A
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	3-5 d	25	F1 Hatching success	F1 eggs	0.062	0.139	0.093	Tapp et al. 1990	В
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	28 d	25	F0 Survival	F0 eggs	0.062	0.139	0.093	Tapp et al. 1990	В
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	56 d	25	F0 Survival	F0 eggs	0.062	0.139	0.093	Tapp et al. 1990	В

Pimephales	Fathead	FT	Meas	97.0%	300 d	25	F0 Egg	F0	0.062	0.139	0.093	Tapp et al.	R
promelas	minnow		Wicas	77.070	300 u	23	Production	eggs	0.002	0.139	0.075	1990	Ь

Reasons for Exclusion

- A. Less sensitive endpoint
- B. Less sensitive life-stage
- C. Test type not preferred (static vs. flow-through)

Table 8. Acute-to-Chronic Ratios used for derivation of the lambda-cyhalothrin chronic criterion.

Species	Common identifier	Test type	Meas/ Nom	Chemical grade	MATC	LC ₅₀	ACR (LC ₅₀ /MATC)	Reference
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.6%	0.31	0.81	2.6129	Hill et al. 1985
Daphnia magna	Daphnid	FT	Meas	94.3%	0.00263	0.013	4.9430	Farrelly & Hamer 1989
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	0.044	0.36	8.1818	Tapp et al. 1990
	= geomean (individu	ıal ACR	(s)				4.73	

Table 9. Excluded chronic toxicity data from studies rated RL, LR, or LL. S: static; SR: static renewal; FT: flow-through. NR: not reported, NC: not calculable.

Species	Common identifier	Test type	Meas/ Nom	Chemical grade	Duration	Temp (°C)	Endpoint	Age/ size	NOEC (μg/L)	LOEC (µg/L)	MATC (μg/L)	Reference	Rating/ Reason
Cyprinodon variegatus	Sheepshead minnow	FT	Meas	96.6%	28 d	25.1	Weight	embryos	0.25	0.38	0.31	Hill et al. 1985	RL 1, 2
Mysidopsis bahia	Mysid	FT	Meas	98.5%	28 d	25	Reproduction (# of young/ female repro. day)	<24 h	0.00022	0.00046	0.00032	Thompson 1987	RL 2, 5
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	3-5 d	25	F0 Hatching success	eggs	≥ 0.273	> 0.273	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	300 d	25	F0 Survival	eggs	≥ 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	28 d	25	F0 Length	eggs	<u>≥</u> 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	56 d	25	F0 Length	eggs	<u>≥</u> 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	300 d	25	F0 Length	eggs	≥ 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	56 d	25	F1 Length	larvae	≥ 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	300 d	25	F0 Weight	eggs	<u>≥</u> 0.139	> 0.139	NC	Tapp et al. 1990	LR 4
Pimephales promelas	Fathead minnow	FT	Meas	97.0%	56 d	25	F1 Weight	larvae	≥ 0.139	> 0.139	NC	Tapp et al. 1990	LR 4

Exclusion Reasons

- 1. Not a standard method
- 2. Saltwater
- 3. Low chemical purity or purity not reported
- 4. Toxicity value not calculable

- 5. Control response not reported
- 6. Low reliability score
- 7. Endpoint not linked to growth, reproduction or survival (Ch. 3, Section 3-2.1.3) 8. Inappropriate test duration (Ch. 3, Section 3-2.1.1)

Table 10. Acceptable multispecies field, semi-field, laboratory, microcosm, mesocosm studies; R= reliable; L= less reliable.

Reference	Habitat	Rating
Farmer et al. 1995	Outdoor artificial mesocosm	R
Gu et al. 2007	Indoor rice paddy-field ecosystem	L
Lauridsen & Friberg 2005	In-stream experimental channels	L
Rasmussen et al. 2008	Outdoor artificial stream channels	L
Roessink et al. 2005	Outdoor artificial ditch microcosm	R
Schroer et al. 2004	Outdoor artificial ditch microcosm	R
Van Wijngaarden et al. 2006	Outdoor artificial ditch microcosm	R
Wendt-Rasch et al. 2004	Outdoor pond microcosms	R

Table 11. Threatened, Endangered, or Rare Species Predicted values by ICE.

Surrogate		Pre	dicted
Species	$LC_{50} (\mu g/L)$	Species	LC_{50} (µg/L)
Rainbow trout (Oncorhynchus mykiss)	0.27	Chinook salmon (O. tshawytscha)	0.539 (0.176-1.65)*
		Coho salmon (O. kisutch)	0.277 (0.180-0.426)
		Lahontan cutthroat trout (O. clarki henshawi)	0.397 (0.197-0.789)*

^{*} Input toxicity value was less than model minimum

Appendix A

Data summary sheets

Abbreviations used in this appendix: NR = Not Reported

Study Ratings:

RR = Relevant, Reliable

RL = Relevant, Less Reliable

LR = Less Relevant, Reliable

LL = Less Relevant, Less Reliable

RN = Relevant, Not Reliable

LN = Less Relevant, Not Reliable

N = Not Relevant

Unused lines deleted from tables

Summary sheets are in alphabetical order according to species

Aedes aegypti

Study: Rodriguez MM, Bisset JA, Fernandez D. 2007. Levels of insecticide resistance and resistance mechanisms in *Aedes aegypti* from some Latin American countries. Journal of the American Mosquito Control Association. 23(4): 420-429.

RelevanceReliabilityScore: 92.5 (No control response)Score: 57Rating: RRating: N

Reference	Rodriguez et al. 2007	A. aegypti
Parameter	Value	Comment
Test method cited	WHO 1981	
Phylum	Arthropoda	
Class	Insecta	
Order	Diptera	
Family	Culicidae	
Genus	Aedes	
Species	aegypti	
Family in North America?	Yes	
Age/size at start of test/growth	Early 4 th instar larvae	
phase		
Source of organisms	Lab culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	24 h	
Data for multiple times?	No	
Effect 1	Survival	
Control response 1	NR	
Temperature	NR	
Test type	Static	
Photoperiod/light intensity	NR	
Dilution water	Tap water	
рН	NR	
Hardness	NR	
Alkalinity	NR	
Conductivity	NR	
Dissolved Oxygen	NR	
Feeding	No	
Purity of test substance	λ-Cyhalothrin: Technical	

Reference	Rodriguez et al. 2007	A. aegypti
Parameter	Value	Comment
Concentrations measured?	No	
Measured is what % of nominal?	NR	
Chemical method documented?	NR	
Concentration of carrier (if any) in	1 mL acetone /100 mL	
test solutions	water	
Concentration 1 Nom/Meas (µg/L)	5 concentrations	20/rep x 2
Control	Water and methanol control	20/rep x 2
λ-Cyhalothrin LC50 (95%	Rockefellar (susceptible): 1	Probit (Finney
Confidence interval) for 8 strains*	(0.8-1)	1971)
in ug/L	Santiago de Cuba: 6 (5-6)	
	Havana City: 30 (20-30)	
	Jamaica: 5 (4-6)	
	Panama: 0.5 (0.4-0.5)	
	Costa Rica: 4 (3-4)	
	Nicaragua: 0.3 (0.3-0.4)	
	Peru: 0.1 (0.1-0.2)	
	Venezuela: 0.6 (0.4-0.7)	

^{*}Rockefellar: laboratory susceptible strain of Caribbean origin, colonized in the early 1930s, provided by the CDC laboratory in San Juan, Puerto Rico.

Santiago de Cuba: natural population collected from Santiago de Cuba, Cuba in 2002 during last dengue epidemic

Havana City: natural population collected from Havana City, Cuba in 2002 during last dengue epidemic

Jamaica: collected in 1998 and maintained in laboratory without exposure to insecticides Costa Rica: collected in 1998 and maintained in laboratory without exposure to insecticides Panama: collected in 1998 and maintained in laboratory without exposure to insecticides Nicaragua: collected in 1998 and maintained in laboratory without exposure to insecticides Peru: collected in 1998 and maintained in laboratory without exposure to insecticides Venezuela: collected in 1998 and maintained in laboratory without exposure to insecticides

Reliability points taken off for:

<u>Documentation:</u> Analytical method (4), Nominal concentrations (3), Measured concentrations (3), Hardness (2), Alkalinity (2), Dissolved Oxygen (4), Temperature (4), Conductivity (2), pH (3), Photoperiod (3), Hypothesis tests (8)

<u>Acceptability:</u> Control response (9), Meas. Concentrations 20% Nom (4), Concentrations not ≥ 2x water solubility (4), Carrier solvent ≤ 0.5 mL/L (4), Organisms randomized (1), Dilution water (2), Hardness (2), Alkalinity (2), Dissolved oxygen (6), Temperature (6), Conductivity (1), pH (2), Photoperiod (2), Test vessels randomized (2), Appropriate spacing between concentrations (2), Hypothesis tests (3)

Asellus aquaticus

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 83.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	A. aquaticus
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Malacostraca	
Order	Isopoda	
Family	Aselloidea	
Genus	Asellus	
Species	aquaticus	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research stations	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	7.4-8.4	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	A. aquaticus
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	7.9-8.3 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 75-126%; 48 h: 56-58%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	0.49/<10/<10	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	1.0/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	2.0/10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	3.9/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	7.8/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	16/12/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 7 Nom/Meas t ₀ /48 h	31/33/18	1 rep, 10 orgs/rep
(ng/L)		
Concentration 8 Nom/Meas t ₀ /48 h	62/78/35	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	26 (18-36) ng/L	Method: Iteratively
		re-weighted linear
		regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Brachydanio rerio Macrobrachium nippoensis

Study: Gu BG, Wang HM, Chen WL, Cai DJ, Shan ZJ. 2007. Risk assessment of lambdacyhalothrin on aquatic organisms in paddy field in China. Regulatory Toxicology and Pharmacology, 48: 69-74.

Relevance

Score: 67.5 (No std method, Low chemical purity, No control response)

Rating: N

Brachydanio rerio

Study: Kent SJ, Shillabeer N. 1997c. Lambda-cyhalothrin: Acute toxicity to zebra danio (*Brachydanio rerio*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Kent & Shillabeer 1997c	B. rerio
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Cypriniformes	
Family	Cyprinidae	
Genus	Brachydanio	
Species	rerio	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	> >11 d old	
	mean control weight	
	and length 0.70 g and	
	36 mm at end of test.	
Source of organisms	Lab culture	Aquatic Research
		Organisms,
		Hampton, NH, USA
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	11 d acclimation in
free?		facility
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	25 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
рН	7.01-7.43	
Hardness	42.3-46.7 mg/L	
Alkalinity	25.6 mg/L	

Reference	Kent & Shillabeer 1997c	B. rerio
Parameter	Value	Comment
Conductivity	207-225 μS/cm	
Dissolved Oxygen	7.4-8.4 mg/L, > 90% sat	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	35-75%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.10/0.035	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.20/0.070	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.40/0.21	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.8/0.40	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	1.6/1.2	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	3.2/1.8	1 rep, 20 org/rep
Control	Solvent and Dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.97 (0.74-1.4)	Method: Moving
(µg/L)	48 h: 0.80 (0.62-1.1)	average angle
	72 h: 0.64 (0.48-0.90)	
	96 h: 0.64 (0.48-0.90)	

LC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Photoperiod (3), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Brachydanio rerio

Study: Wang W, Cai DJ, Shan ZJ, Chen WL, Poletika N, Gao XW. 2007. Comparison of the acute toxicity for gamma-cyhalothrin and lambda-cyhalothrin to zebra fish and shrimp. Regulatory Toxicology and Pharmacology, 47: 184-188.

RelevanceReliabilityScore: 75 (No standard method, low chemical purity)Score: 61Rating: LRating: L

Reference	Wang et al. 2007	B. rerio
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Chordata	
Class	Osteichthyes	
Order	Cypriniformes	
Family	Cyprinidae	
Genus	Brachydanio	
Species	rerio	
Family in North America?	Yes	
Age/size at start of test/growth phase	30-45 d old, 0.38 g, 3.5 cm	
Source of organisms	Lab culture	Nanjing Institute of Environmental Sciences
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes, 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	25 ± 2 °C	
Test type	Static renewal, renewed	
	every 24 h	
Photoperiod/light intensity	NR	
Dilution water	NR	
pH	7.1	
Hardness	6.8-8.0 °HG	
Alkalinity	NR	
Conductivity	NR	

Reference	Wang et al. 2007	B. rerio
Parameter	Value	Comment
Dissolved Oxygen	NR	
Feeding	None during test	
Purity of test substance	Kung Fu 25 EW formulation	
Concentrations measured?	Yes, but NR	
Measured is what % of nominal?	NR	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	None used	
test solutions		
Concentration 1 Nom/Meas (a.i.	0.5	1 rep, 10 orgs/rep
μg/L)		
Concentration 2 Nom/Meas (a.i.	1.0	1 rep, 10 orgs/rep
μg/L)		
Concentration 3 Nom/Meas (a.i.	2.0	1 rep, 10 orgs/rep
μg/L)		
Concentration 4 Nom/Meas (a.i.	4.0	1 rep, 10 orgs/rep
μg/L)		
Concentration 5 Nom/Meas (a.i.	6.0	1 rep, 10 orgs/rep
μg/L)		
Concentration 6 Nom/Meas (a.i.	10	1 rep, 10 orgs/rep
μg/L)		
Concentration 7 Nom/Meas (a.i.	20	1 rep, 10 orgs/rep
μg/L)		
Control	Dilution water	1 rep, 10 orgs/rep
LC ₅₀ (95% confidence interval)	24 h: 8.26 (5.93-11.51)	Method: NR
(μg/L)	48 h: 3.91 (2.62-5.84)	
	72 h: 2.05 (1.40-3.01)	
	96 h: 1.94 (1.33-2.84)	

LC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Chemical purity (5), Measured concentrations (3), Dilution water (3), Alkalinity (2), Dissolved oxygen (4), Conductivity (2), Photoperiod (3), Statistical methods (5), Hypothesis tests (8),

Acceptability: Standard method (5), Chemical purity (10), Measured concentrations w/in 20% nominal (4), Concentrations > 2x solubility (4), Organisms randomized (1), Alkalinity (2), Dissolved oxygen (6), Conductivity (1), Photoperiod (2), Random design (2), Adequate replication (2), Statistical method (2), Hypothesis tests (3).

Caridina laevis

Study: Sucahyo D, van Straalen NM, Krave A, van Gestel CAM. 2008. Acute toxicity of pesticides to the tropical freshwater shrimp *Caridina laevis*. Ecotoxicology and Environmental Safety, 69: 421-427.

RelevanceReliabilityScore: 75 (No standard method, Low chemical purity)Score: 73Rating: LRating: R

Reference	Sucahyo et al. 2008	C. laevis
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Arthropoda	
Class	Malacostraca	
Order	Decapoda	
Family	Atyidae	
Genus	Caridina	
Species	laevis	
Family in North America?	Yes	
Age/size at start of test/growth	Adults, 15-20 mm	
phase		
Source of organisms	Freshwater lake, Indonesia	
Have organisms been exposed to	Possibly	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes, 24 h	
Effect 1	Mortality	
Control response 1	< 10%	
Temperature	26 ± 1 °C	
Test type	Static	
Photoperiod/light intensity	12 L: 12 D	
Dilution water	Dechlorinated tapwater	
рН	6.9-7.2	
Hardness	128-136 mg/L	
Alkalinity	NR	
Conductivity	NR	
Dissolved Oxygen	6.8-7. 2 mg/L	
Feeding	None during test	
Purity of test substance	25 g/L formulation	

Reference	Sucahyo et al. 2008	C. laevis
Parameter	Value	Comment
Concentrations measured?	No	
Measured is what % of nominal?	NR	
Chemical method documented?	No	
Concentration of carrier (if any) in	None used	
test solutions		
Concentration 1 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Concentration 2 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Concentration 3 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Concentration 4 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Concentration 5 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Concentration 6 Nom/Meas (µg/L)	NR	5 reps, 20 orgs/rep
Control	Dilution water	5 reps, 20 orgs/rep
LC ₅₀ (95% confidence interval)	24 h: 0.87 (0.76-0.98)	Method: trimmed
$(\mu g/L)$	96 h: 0.33 (0.30-0.37)	Spearman-Karber
NOEC (µg/L)	0.1	Method: Tukey's
		test
		p: NR
		MSD: NR
LOEC (µg/L)	0.2	Same as above
MATC (GeoMean NOEC,LOEC)	0.14 μg/L	

LC₅₀ or NOEC/LOEC calculated based on nominal active ingredient concentrations.

Reliability points taken off for:

<u>Documentation:</u> Analytical method (4), Nominal concentrations (3), Measured concentration (3), Alkalinity (2), Conductivity (2), Hypothesis tests (8). <u>Acceptability:</u> Standard method (5), Measured concentrations w/in 20% nominal (4), Prior contamination (4), Exposure type appropriate (2), Alkalinity (2), Conductivity (1), Random

design (2), Hypothesis tests (3).

Ceriodaphnia dubia

Study: Wheelock CE, Miller JL, Miller MJ, Gee SJ, Shan G, Hammock BD. 2004. Development of toxicity identification evaluation procedures for pyrethroid detection using esterase activity. Environmental Toxicology and Chemistry 23(11): 2699-2708

RelevanceReliabilityScore: 100Score: 74Rating: RRating: R

Reference	Wheelock et al. 2004	C. dubia
Parameter	Value	Comment
Test method cited	EPA	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Ceriodaphnia	
Species	dubia	
Family in North America?	Yes	
Age/size at start of test/growth	< 24 h	
phase		
Source of organisms	Lab culture; AQUA-	
	Science, Davis, CA	
Have organisms been exposed to	Probably not	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	Yes	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Survival	
Control response 1	> 90%	
Temperature	25 +/- 1 °C	
Test type	Static	
Photoperiod/light intensity	16:8 light: dark	
Dilution water	EPA moderately hard	
рН	7.4-7.8	
Hardness	80-100 mg/L	
Alkalinity	60-70 mg/L	
Conductivity	Measured but NR	
Dissolved Oxygen	Measured but NR	
Feeding	None during test	

Reference	Wheelock et al. 2004	C. dubia
Parameter	Value	Comment
Purity of test substance	>97%	
Concentrations measured?	No	
Measured is what % of nominal?	NR	
Chemical method documented?	NR	
Concentration of carrier (if any) in	<0.1%	
test solutions		
Concentration 1 Nom/Meas (µg/L)	5-7 concentrations	2-4 w/ 5 neonates
		each, distributed in
		'stratified random
		assortment'
Control	Water and methanol	2-4 w/ 5 neonates
	control	each
LC50; indicate calculation method	48 h: 0.200 +/- 0.090 ug/L	ToxCal software, but
		no stat method
		reported

Reliability points taken off for:

<u>Documentation:</u> Nominal concentrations (3), Measured concentrations (3), Dissolved Oxygen (4), Conductivity (2), Statistical methods identified (5), Hypothesis tests (8)

<u>Acceptability:</u> Meas. Concentrations 20% Nom (4), Carrier solvent \leq 0.5 mL/L (4), Exposure type (2), Appropriate spacing between concentrations (2), Appropriate statistical method (2), Hypothesis tests (3)

Ceriodaphnia dubia Daphnia magna

Study: Mokry, LE & Hoagland KD. 1990. Acute toxicities of five synthetic pyrethroid insecticides to *Daphnia magna* and *Ceriodaphnia dubia*. Environmental Toxicology & Chemistry 9 (8): 1045-1051.

Relevance

Score: 67.5 (purity-25.4 %, no std method, control response NR)

Rating: N

Channa punctatus

Study: Kumar A, Sharma B, Pandey RS. 2007. Preliminary evaluation of the acute toxicity of cypermethrin and lambda-cyhalothrin to *Channa punctatus*. Bull Environ Contam Toxicol, 79: 613-616.

RelevanceReliabilityScore: 75 (No std method, Low chemical purity)Score: 67Rating: LRating: L

Reference	Kumar et al. 2007	C. punctatus
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Chordata	
Class	Actinopterygii	
Order	Perciformes	
Family	Channidae	
Genus	Channa	
Species	punctatus	
Family in North America?	Not native, but is an invasive	
-	species	
Age/size at start of test/growth	Teleosts, 11-13 cm, 23 ± 2 g	
phase		
Source of organisms	Local fish market in India	
Have organisms been exposed to	Possibly	
contaminants?		
Animals acclimated and disease-	Yes, 2 week acclimation	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	No	
Effect 1	Mortality	
Control response 1	0%	
Temperature	27 ± 1 °C	
Test type	SR, 24 h renewal	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
pН	6.8 ± 2 °C	
Hardness	$113.3 \pm 2 \text{ mg/L}$	
Alkalinity	NR	
Conductivity	NR	
Dissolved Oxygen	$6.9 \pm 0.4 \text{ mg/L}$	
Feeding	None	

Reference	Kumar et al. 2007	C. punctatus
Parameter	Value	Comment
Purity of test substance	5%	
Concentrations measured?	No	
Measured is what % of nominal?	NR	
Chemical method documented?	NR	
Concentration of carrier (if any) in	NR, acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	2.5	1 rep, 12 org/rep
Concentration 2 Nom/Meas (µg/L)	5	1 rep, 12 org/rep
Concentration 3 Nom/Meas (µg/L)	7.5	1 rep, 12 org/rep
Concentration 4 Nom/Meas (µg/L)	10	1 rep, 12 org/rep
Concentration 5 Nom/Meas (µg/L)	12.5	1 rep, 12 org/rep
Concentration 6 Nom/Meas (µg/L)	15	1 rep, 12 org/rep
Control	Solvent	1 rep, 12 org/rep
LC ₅₀ (95% confidence interval)	7.92	Method: Karber
(µg/L)		arithmetic method

LC₅₀ calculated based on nominal concentrations.

The three highest concentrations tested are $\geq 2x$ the water solubility of lambda-cyhalothrin. Behavioral effects were also observed.

Reliability points taken off for:

<u>Documentation:</u> Analytical method (4), Measured concentrations (3), Alkalinity (2), Conductivity (2), Photoperiod (3), Hypothesis tests (8).

Acceptability: Standard method (5), Chemical purity (10), Measured concentrations w/in 20% of nominal (4), Concentrations exceed 2x solubility (4), Carrier solvent concentration (4), Prior contamination of organism (4), Organisms randomized (1), Alkalinity (2), Conductivity (1), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Channa punctatus

Study: Kumar A, Sharma B, Pandey RS. 2008. Cypermethrin and l-cyhalothrin induced alterations in nucleic acids and protein contents in a freshwater fish, *Channa punctatus*. Fish Physiol Biochem, 34:331-338.

RelevanceReliabilityScore: 45*Score: not ratedRating: NRating:

^{*}No standard method, Endpoint not linked to survival/growth/reproduction, Low chemical purity, No calculable toxicity values.

Channa punctatus (Bloch)

Study: Kumar A, Rai DK, Sharma B, Pandey RS. 2009. λ-cyhalothrin and cypermethrin induced *in vivo* alterations in the activity of acetylcholinesterase in a freshwater fish, *Channa punctatus* (Bloch). Pesticide Biochemistry and Physiology, 93:96-99.

RelevanceReliabilityScore: NScore: not ratedRating: 45*Rating:

^{*}No standard method, Endpoint not linked to survival/growth/reproduction, Low chemical purity, No calculable toxicity values.

Chaoborus sp.

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	Chaoborus sp.
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Insecta	
Order	Diptera	
Family	Chaoboridae	
Genus	Chaoborus	
Species	NR	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research station	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	7.4-8.4	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	Chaoborus sp.
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.0-8.8 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 100-113%; 48 h: 52-63%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	0.49/<10/<10	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	1.0/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	2.0/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	3.9/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	7.8/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	16/16/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 7 Nom/Meas t ₀ /48 h	31/35/16	1 rep, 10 orgs/rep
(ng/L)	(2)/70/20	1 10 /
Concentration 8 Nom/Meas t ₀ /48 h	62/70/39	1 rep, 10 orgs/rep
(ng/L)	Dil di di di	1 10 /
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	2.8 (1.8-4.1) ng/L	Method: Iteratively
		re-weighted linear
		regression

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), No prior contamination exposure (4), Organisms randomized (1), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Cloeon dipterum

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	C. dipterum
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Insecta	
Order	Ephemeroptera	
Family	Baetidae	
Genus	Cloeon	
Species	dipterum	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research stations	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	8.2-8.7	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	C. dipterum
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.6-9.0 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 94-116%; 48 h: 45-47%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	0.49/<10/<10	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	1.0/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	2.0/10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	3.9/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	7.8/<10/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	16/16/<10	1 rep, 10 orgs/rep
(ng/L)		
Concentration 7 Nom/Meas t ₀ /48 h	31/29/14	1 rep, 10 orgs/rep
(ng/L)		
Concentration 8 Nom/Meas t ₀ /48 h	62/72/29	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	38 (23-93) ng/L	Method: Iteratively
		re-weighted linear
		regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Prior contamination (4), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random

design (2), Appropriate statistical method (2), Hypothesis tests (3).

Corixa sp.

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	Corixa sp.
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Insecta	
Order	Hemiptera	
Family	Corixidae	
Genus	Corixa	
Species	NR	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research station	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	Solvent: 10%	
Effect 2	Mortality	
Control response 2	Solvent: 10%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	8.4-8.6	
Hardness	179 mg/L	
Alkalinity	150 m6g/L	

Reference	Hamer <i>et al.</i> 1998	Corixa sp.
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.4-9.1 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 114-125%; 48 h: 50-75%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	16/20/12	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	31/37/20	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	62/74/31	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	125/143/67	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	30 (21-42) ng/L	Method: Iteratively
		re-weighted linear
		regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8) <u>Acceptability:</u> No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Prior contaminant exposure (4), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Crassostrea gigas

Study: Thompson RS. 1985. PP321: Determination of the acute toxicity to larvae of the Pacific oyster (*Crassostrea gigas*). ICI Agrochemicals. DPR study 50907-087.

Relevance

Score: n/a

Rating: N \rightarrow because all concentrations tested were > 2x solubility

Culex quinquefasciatus

Study: Halliday WR Georghiou GP. 1985. Cross-resistance and dominance relationships of pyrethroids in a permethrin-selected strain of Culex quinquefasciatus (Diptera: Culicidae). Journal of Economic Entomology 78: 127-1232.

RelevanceReliabilityScore: 82.5 (No std method, Control not described)Score: 47Rating: LRating: N

Reference	Halliday & Georghiou 1985	<i>C</i> .
		quinquefasciatus
Parameter	Value	Comment
Test method cited	Ref Georghiou 1966	
Phylum	Arthropoda	
Class	Insecta	
Order	Diptera	
Family	Culicidae	
Genus	Culex	
Species	quinquefasciatus	
Family in North America?	Yes	
Age/size at start of test/growth	4 th instar	
phase		
Source of organisms	Lab culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	24 h	
Data for multiple times?	No	
Effect 1	Mortality	Susceptible and
		resistant strains
		tested
Control response 1	< or = 15%	
Temperature	NR	
Test type	static	
Photoperiod/light intensity	NR	
Dilution water	tap	
pH	NR	
Hardness	NR	
Alkalinity	NR	

Reference	Halliday & Georghiou 1985	C. quinquefasciatus
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	NR	
Feeding	NR	
Purity of test substance	'Technical' no%	
Concentrations measured?	NR	
Measured is what % of nominal?	NR	
Chemical method documented?	NR	
Concentration of carrier (if any) in	10 mL/L	
test solutions		
Concentration 1 Nom/Meas (µg/L)	4 levels, but concentrations	4 reps and 20
	not reported	organisms per rep
Control	yes	
LC50; indicate calculation method	0.73 ug/L - susceptible	probit
	220 ug/L - resistant	

Reliability points taken off for:

<u>Documentation:</u> Control Type (8), Analytical method (4), Nominal concentrations (3), Measured concentrations (3), Hardness (2), Alkalinity (2), Dissolved Oxygen (4), Temperature (4), Conductivity (2), pH (3), Photoperiod (3) Hypothesis tests (8)

Acceptability: Standard method (5), Control appropriate type (6), Meas. Concentrations 20% Nom (4), Concentrations do not exceed 2x water solubility (4), Carrier solvent ≤ 0.5 mL/L (4), Appropriate age/ size (3), Organisms randomly assigned to containers (1), Dilution water (2), Hardness (2), Alkalinity (2), Dissolved Oxygen (6), Temperature (6), Conductivity (1), pH (2), Photoperiod (2), Adequate number of concentrations (3), Appropriate spacing between concentrations (2), Random / block design (2), Hypothesis tests (3)

Cyclops sp.

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	Cyclops sp.
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Maxillopoda	
Order	Cyclopoida	
Family	Cyclopidae	
Genus	Cyclops	
Species	NR	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research station	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	Dil Water: 0%	
	Solvent: 20%	
Effect 2	Mortality	
Control response 2	Dil Water: 0%	
	Solvent: 20%	
Temperature	$20 \pm 2^{\circ}\text{C}$	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
pH	8.1-8.7	

Reference	Hamer <i>et al.</i> 1998	Cyclops sp.
Parameter	Value	Comment
Hardness	179 mg/L	
Alkalinity	150 mg/L	
Conductivity	NR	
Dissolved Oxygen	7.9-8.9 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 83-109%; 48 h: 35-53%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	62/65/29	1 rep, 10 orgs/rep
(µg/L)		
Concentration 2 Nom/Meas t ₀ /48 h	125/117/57	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	250/207/88	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	500/485/266	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	1000/1031/419	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	2000/2184/726	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	300 (200-460) ng/L	Method: Iteratively
		re-weighted linear
		regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8) <u>Acceptability:</u> No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Prior contaminant exposure (4), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Cyprinodon variegatus

Study: Hill RW. 1985. PP321: Determination of acute toxicity to sheepshead minnow (*Cyprinodon variegatus*). ICI Agrochemicals. DPR Study 50907-085.

RelevanceReliabilityScore: 85 (saltwater)Score: 77Rating: LRating: R

Reference	Hill 1985	C. variegatus
Parameter	Value	Comment
Test method cited	USEPA 1982	
Phylum	Chordata	
Class	Actinopterygii	
Order	Cyprinodontiformes	
Family	Cyprinodontidae	
Genus	Cyprinodon	
Species	variegatus	
Family in North America?	Yes	
Age/size at start of test/growth	0.60 g, 27.4 mm	
phase		
Source of organisms	Commercial lab	Sea Plantations, Inc.
		Salem MA
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	17 ± 1°C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Seawater from Torbay,	
	Devon, UK	
рН	8.0-8.1	
Hardness	NR	
Alkalinity	NR	
Salinity	34.97 o/oo	
Dissolved Oxygen	6.2-6.8 mg/L, >82% sat	
Feeding	NR	

Reference	Hill 1985	C. variegatus
Parameter	Value	Comment
Purity of test substance	96.5%	
Concentrations measured?	Yes	
Measured is what % of nominal?	51.8-75%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	16 mg/L acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.56/0.29	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	1.0/0.55	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	1.8/1.35	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	2.4/1.72	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	3.2/2.37	1 rep, 20 org/rep
Control	Solvent and dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 1.34	Method: Probit
$(\mu g/L)$	48 h: 1.14	
	72 h: 0.85	
	96 h: 0.81	

LC₅₀ calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hardness (2), Alkalinity (2), Conductivity (2), Photoperiod (3), Hypothesis tests (8).

<u>Acceptability:</u> Carrier solvent (4), Organisms randomized (1), Feeding (3), Acceptable dilution water (2), Hardness (2), Alkalinity (2), Conductivity (1), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Cyprinodon variegatus

Study: Hill RW, Caunter JE, Cumming RI. 1985. PP321: Determination of the chronic toxicity to sheepshead minnow (*Cyprinodon variegatus*) embryos and larvae. DPR study number 50907-088.

RelevanceReliabilityScore: 75 (No standard method, saltwater)Score: 81Rating: LRating: R

Reference	Hill et al. 1985	C. variegatus
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Chordata	
Class	Actinopterygii	
Order	Cyprinodontiformes	
Family	Cyprinodontidae	
Genus	Cyprinodon	
Species	variegatus	
Family in North America?	Yes	
Age/size at start of test/growth	Embryos and larvae (just	
phase	hatched – 28 d posthatch)	
Source of organisms	Lab stock culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	Yes	
Test duration	28 d	
Data for multiple times?	No	
Effect 1	Embryo % hatch	
Control response 1	Solvent cont: 91.6%	
	Dil water:88.6%	
Effect 2	Length at 28 d post-hatch	
Control response 2	Solvent cont: 18.4 mm	
	Dil water: 18.4 mm	
Effect 3	Weight at 28 d post-hatch	
Control response 3	Solvent cont: 181.7 mg	
	Dil water: 172 mg	
Effect 4	28 d Survival (from initial	
	embryos)	
Control response 4	Solvent: 83.1%	
	Dil water: 85.3%	

Reference	Hill et al. 1985	C. variegatus
Parameter	Value	Comment
Effect 5	28 d Survival (from hatched	
	embryos only)	
Control response 5	Solvent: 90.8%	
	Dil water: 96.5%	
Temperature	$25.1 \pm 1^{\circ}$ C	
Test type	Flow-through	
Photoperiod/light intensity	12 L: 12 D, 2800-3300 lux	
Dilution water	Filtered seawater mixed with	
	freshwater	
рН	8.2-8.3	
Hardness	NR	
Alkalinity	NR	
Salinity	23.5-26.7 o/oo	
Dissolved Oxygen	6.0-7.6 mg/L	
Feeding	2-3x daily	
Purity of test substance	96.6%	
Concentrations measured?	Yes	
Measured is what % of nominal?	Mean: 41%	Range: 36-46.9%
Chemical method documented?	Yes, GC	
Concentration of carrier (if any) in	NR	
test solutions		
Concentration 1 Nom/Meas (µg/L)	1.0/0.38	2 reps, 30 orgs/rep
Concentration 2 Nom/Meas (µg/L)	0.56/0.25	2 reps, 30 orgs/rep
Concentration 3 Nom/Meas (µg/L)	0.32/0.14	2 reps, 30 orgs/rep
Concentration 4 Nom/Meas (µg/L)	0.18/0.07	2 reps, 30 orgs/rep
Concentration 5 Nom/Meas (µg/L)	0.010/0.04	2 reps, 30 orgs/rep
Control	Solvent and Dil. water	2 reps, 30 orgs/rep
NOEC (μg/L)	Weight: 0.25	Method: 1 way
		ANOVA and
		Dunnett's test
		p: 0.05 (and 0.01)
		MSD: NR
LOEC (µg/L)	Weight: 0.38	Same as above
MATC (GeoMean NOEC,LOEC)	Weight: 0.31 µg/L	
% control at NOEC	Weight: 99.0%	Solvent control
		used in calculation
% of control LOEC	Weight: 86.8%	Solvent control
		used in calculation

Weight was the only endpoint that was significantly affected at any concentration and the NOEC/LOEC were calculated based on weight data only.

NOEC/LOEC calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hardness (2), Alkalinity (2), Conductivity (2), Hypothesis tests (4), Point estimates (8)

Acceptability: Standard method (5), Measured concentrations w/in 20% of nominal (4), Carrier solvent % (4), Hardness (2), Alkalinity (2), Conductivity (1), Adequate replication (2), Minimum significant difference (1), Point estimates (3).

Danio rerio

Study: Xu C, Wang J, Liu W, Sheng GD, Tu Y, Ma Y. 2008. Separation and aquatic toxicity of enantiomers of the pyrethroid insecticide lambda-cyhalothrin. Environmental Toxicology and Chemistry, 27: 174-181.

Relevance Reliability

Score: Fish – 85 (No control info)

Rating: Fish – L

Score: Fish – 56

Rating: Fish – N

Eggs – N (all concentrations tested $\geq 2x$ water solubility)

Daphnia magna

Study: Barata C, Baird DJ, Nogueira AJA, Soares AMVM, Riva MC. 2006. Toxicity of binary mixtures of metals and pyrethroid insecticides to *Daphnia magna* Straus. Implications for multi-substance risks assessment. Aquatic Toxicology 78: 1-14.

RelevanceReliabilityScore: A: 100, C: 60Score: A: 78.5Rating: A: R, C: NRating: A: R

C: No std method, Endpoint, Toxicity value

Reference	Barata et al. 2006	D. magna
Parameter	Value	Comment
Test method cited	A: OECD, C: None	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth phase	4 th instar juveniles	
Source of organisms	Lab culture	
Have organisms been exposed to contaminants?	Probably not	
Animals acclimated and disease-free?	Yes	
Animals randomized?	A: NR, C: Yes	
Test vessels randomized?	NR	
Test duration	A: 48 h, C: 24 h	
Data for multiple times?	No	
Effect 1	A: Immobility	
Control response 1	100% survival	
Effect 2	C: Feeding rate	
Control response 2	Dil. $5.25 \pm 0.38 \times 10^5$	Sol: $5.27 \pm 0.54 \text{ x}$
	cells/ind/h	10 ⁵ cells/ind/h
Temperature	NR	
Test type	Static	
Photoperiod/light intensity	A: NR, C: 24h dark	
Dilution water	ASTM hard synthetic water	
рН	8.3 ± 0.2	
Hardness	NR	
Alkalinity	NR	

Reference	Barata et al. 2006	D. magna
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	≥ 91%	
Feeding	A: None during test, C: yes	
Purity of test substance	99%	
Concentrations measured?	Yes	
Measured is what % of nominal?	Mean 85.5% (Table 2)	
Chemical method documented?	Ref. McWilliam & Baird 2002	
Concentration of carrier (if any) in test solutions	<0.5% acetone	
Concentration 1 Nom (nmol/L)	A: 0.15 (Fig 2)	A: 3 x 10/conc C: 5 x 5/conc
Concentration 2 Nom (nmol /L)	A: 0.25 (Fig 2)	A: 3 x 10/conc C: 5 x 5/conc
Concentration 3 Nom (nmol /L)	A: 0.56	A: 3 x 10/conc
Meas	0 h: 0.43 (0.09), 48 h: 0.23 (0.06)	C: 5 x 5/conc
Concentration 4 Nom (nmol /L)	A: 0.8 (Fig 2)	A: 3 x 10/conc
	A 1 (E: 2)	C: 5 x 5/conc
Concentration 5 Nom (nmol/L)	A: 1 (Fig 2)	A: 3 x 10/conc
Meas	A 1.22	C: 5 x 5/conc
Concentration 6 Nom (nmol /L)	A: 1.33	A: 3 x 10/conc
Meas	0h: 1.03 (0.11), 48 h: 0.52 (0.08)	C: 5 x 5/conc
Concentration 7 Nom (nmol /L)	A: 2.22	A: 3 x 10/conc
Meas	0 h: 1.73 (0.18), 48 h: 0.83 (0.13)	C: 5 x 5/conc
Control	Solvent control	A: 3 x 10/conc
		C: 5 x 5/conc
EC50; indicate calculation method	A: 0.87 (0.86-0.88) nmol/L	A: linear regression,
	0.39 ug/L	p<0.05, calc. w/
	C: 0.22 (0.21-0.23) nmol/L	meas conc.
	0.10 ug/L	C: p<0.05

Reliability points taken off for:

<u>Acute Documentation:</u> Hardness (2), Alkalinity (2), Temperature (4), Conductivity (2), Photoperiod (3), Hypothesis tests (8)

<u>Acute Acceptability:</u> Meas. Concentrations 20% Nom (4), Organisms randomly assigned (1), Hardness (2), Alkalinity (2), Temperature (6), Conductivity (1), Photoperiod (2), Test vessels randomized (2), Hypothesis tests (3)

Daphnia magna

Study: Barata C, Baird DJ, Nogueira AJA, Soares AMVM, Riva MC. 2007. Life-history responses of *Daphnia magna* Straus to binary mixtures of toxic substances: Pharmacological versus ecotoxicological modes of action. Aquatic Toxicology 84: 439-449.

RelevanceReliabilityScore: 90 (No std method)Score: 80Rating: RRating: R

Reference	Barata et al. 2007	D. magna
Parameter	Value	Comment
Test method cited	None	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth phase	Egg production: < 24 h neonates, exposed until 8-9 d old, when egg production of 2 nd and 3 rd clutches began (1 st clutch not measured because they were not exposed for entire lifetime) Feeding: female adults (after 2 nd brood, to avoid molting)	
Source of organisms	Lab culture	
Have organisms been exposed to contaminants?	Probably not	
Animals acclimated and disease-free?	Yes	
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	Egg production: 6 d Feeding: 24 h	
Data for multiple times?	no	
Effect 1	Egg production	
Control response 1	Exp 1: 52.2 ± 2.6 eggs/female Exp 2: 37.1 ± 7.7 eggs/female	
Effect 2	Feeding rate	
Control response 2	Exp 1: $10.27 \pm 0.11 \times 10^5$ cells/ind/h	

Reference	Barata et al. 2007	D. magna
Parameter	Value	Comment
	Exp 2: $8.67 \pm 1.28 \times 10^5$	
	cells/ind/h	
Temperature	20 ± 1°C	
Test type	NR, probably Static	
Photoperiod/light intensity	14 L: 10 D	
Dilution water	ASTM hard synthetic water	
pН	8.3 ± 0.2	
Hardness	NR	
Alkalinity	NR	
Conductivity	NR	
Dissolved Oxygen	≥ 91%	
Feeding	Yes with algae (<i>C. vulgaris</i>)	
Purity of test substance	99%	
Concentrations measured?	Yes – only 3 highest	
Measured is what % of nominal?	~60%	
Chemical method documented?	Yes, HPLC	
Concentration of carrier (if any) in	< 0.05% acetone	
test solutions		
Concentration 1 Nom (nmol/L)	0.1	
Concentration 2 Nom/t ₀ Meas/24 h	$0.31/0.22 \pm 0.04/0.16 \pm 0.07$	
Meas (nmol/L)		
Concentration 3 Nom/t ₀ Meas/24 h	$0.56/0.39 \pm 0.07/0.28 \pm 0.07$	
Meas (nmol/L)		
Concentration 4 Nom/t ₀ Meas/24 h	$0.9/0.62 \pm 0.07/0.46 \pm 0.09$	
Meas (nmol/L)		
Control	Solvent control	
EC ₅₀ (95% confidence interval)	Feeding: 0.27 (0.15-0.39)	Method: nonlinear
	nmol/L	allosteric decay
	0.12 ug/L	regression and least
	Egg production: 0.43 (0.39-	squares
	0.47) nmol/L	
	0.2 ug/L	

Point estimates calculated with measured concentrations.

Chronic EC50 values do not appear in data tables because they could not be incorporated into criteria derivation.

Reliability points taken off for:

<u>Documentation:</u> Exposure type (5), Hardness (2), Alkalinity (2), Conductivity (2), Hypothesis tests (8)

<u>Acceptability:</u> Acceptable standard (5), Measured concentrations w/in 20% of nominals (4), Adequate #/rep (2), Exposure type (2), Conductivity (1), Test vessels randomized (2), Statistical method (2), Hypothesis tests (3).

Daphnia magna

Study: Farrelly E, Hamer MJ. 1989. PP321: *Daphnia magna* life-cycle study using a flow-through system. ICI Agrochemicals. MRID 41217501.

RelevanceReliabilityScore: 100Score: 90.5Rating: RRating: R

Reference	Farrelly & Hamer 1989	D. magna
Parameter	Value	Comment
Test method cited	USEPA 1986	EPA 540/9-86-141
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth	Adults < 24 hr old	
phase		
Source of organisms	Lab culture	Jealott's Hill
		facility
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	Yes	
Test duration	21 d	
Data for multiple times?	Yes	3, 7, 14, 21 d
Effect 1	Survival	
Control response 1	3, 7, 14 d: 100% survival	
	21 d: 80% (solvent), 60%	
	(Dil water)	
Effect 2	Growth (length)	
Control response 2	3.48 mm (solvent),	
	3.51 mm (Dil water)	
Effect 3	Reproduction	Total young
		produced & # of
		young/female/day
Control response 3	Total Young: 67.7 (solvent),	
-	78.9 (Dil water)	
	Young/female/d: 5.5	
	(solvent), 6.1 (Dil water)	

Reference	Farrelly & Hamer 1989	D. magna
Parameter	Value	Comment
Temperature	$20 \pm 2^{\circ}\text{C}$	
Test type	FT	
Photoperiod/light intensity	16L:8d, 1200 lux	
Dilution water	Hard reconstituted water	Salts added to DI water
рН	8.1-8.2	Meas. at 0, 7, 14, 21 d
Hardness	165-175 mg/L	
Alkalinity	115-125 mg/L	
Conductivity	555-590 uS/cm	
Dissolved Oxygen	> 89% sat (> 8.2 mg/L)	Meas. at 0, 7, 14, 21 d
Feeding	Yes, 2x/d	0.25 ml Chlorella vulgaris & 0.25 ml active dried yeast
Purity of test substance	94.3%	
Concentrations measured?	Yes	
Measured is what % of nominal?	48-81%	
Chemical method documented?	Yes; LSC & HPLC	meas 1x/wk
Concentration of carrier (if any) in test solutions	NR %	Triethylene glycol
Concentration 1 Nom/Meas (µg/L)	1.024/0.83	Growth/Repro: 7 reps, 1 org/rep Survival: 3 reps, 5 orgs/rep
Concentration 2 Nom/Meas (ng/L)	2.56/1.98	Same as above
Concentration 3 Nom/Meas (ng/L)	6.4/3.50	Same as above
Concentration 4 Nom/Meas (ng/L)	16/9.37	Same as above
Concentration 5 Nom/Meas (ng/L)	40/19.1	Same as above
Control	Solvent and Dil water	Same as above
LC ₅₀ (95% CI, if calculable)	3d: 13 ng/L (10-17) 7d: 8.3 ng/L 14d: 6.9 ng/L (5.3-8.9) 21 d: 3.6 ng/L*	Method: Probit
NOEC	Repro: 1.98 ng/L Growth: 9.27 ng/L	Method: ANOVA p: 0.05 MSD: NR
LOEC	Repro: 3.5 ng/L	Same as above
MATC (GeoMean NOEC,LOEC)	Repro: 2.63 ng/L	
% control at NOEC	NR	
% of control LOEC	NR	

*unacceptable control response at 21 d for survival

- □ LC₅₀ values calculated with measured concentrations
- □ In the flow-through system, the pumps were not pumping the set uL/d, so the nominal concentrations are not representative.
- □ Some isomerization was observed by day 21 of the study.

Reliability points taken off for:

<u>Documentation:</u> Hypothesis tests (6)

<u>Acceptability:</u> Measured conc w/in 20% of nominal (4), Carrier solvent (4), Organisms randomized (1), Temperature variance (3), Minimum significant difference (1)

Daphnia magna

Study: Farrelly E, Hamer MJ, Hill IR. 1984. PP321: Toxicity to first instar *Daphnia magna*. DPR study number 50907-008. ICI Agrochemicals, Plant Protection Division.

RelevanceReliabilityScore: 100Score: 86Rating: RRating: R

Reference	Farrelly et al. 1984	D. magna
Parameter	Value	Comment
Test method cited	USEPA, OECD, ASTM	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth phase	< 24 h, 1 st instar	
Source of organisms	Continuous lab culture at test facility	
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	Yes; 24 h	
Effect 1	Immobility	
Control response 1	0%	
Temperature	20 ± 1 °C	
Test type	Static	
Photoperiod/light intensity	16 L: 8 D	
Dilution water	EPA reconstituted hard water	
рН	8.0-8.6	
Hardness	NR	
Alkalinity	NR	
Conductivity	NR	
Dissolved Oxygen	> 7.6 mg/L, >82% sat	
Feeding	None during test	
Purity of test substance	96.5%	

Reference	Farrelly et al. 1984	D. magna
Parameter	Value	Comment
Concentrations measured?	Yes	
Measured is what % of nominal?	51-68%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	32/17.2	2 tests, 3 reps/test,
		10 orgs/rep
Concentration 2 Nom/Meas (µg/L)	16/8.4	Same as above
Concentration 3 Nom/Meas (µg/L)	8.0/4.1	Same as above
Concentration 4 Nom/Meas (µg/L)	4.0/2.3	Same as above
Concentration 5 Nom/Meas (µg/L)	2.0/1.03	Same as above
Concentration 6 Nom/Meas (µg/L)	1.0/0.52	Same as above
Concentration 7 Nom/Meas (µg/L)	0.5/0.27	Same as above
Concentration 8 Nom/Meas (µg/L)	0.25/0.17	Same as above
Concentration 9 Nom/Meas (µg/L)	0.125/0.08	Same as above
Concentration 10 Nom/Meas (µg/L)	0.0625/0.04 (Test 2 only)	Same as above
Control	Solvent	Same as above
EC ₅₀ (95% confidence interval)	24 h: 5.04	Method: Weighted
(μg/L)	48 h: 0.36	linear regression

EC₅₀ calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hardness (2), Alkalinity (2), Conductivity (2), Hypothesis tests (8) <u>Acceptability:</u> Measured concentrations w/in 20% of nominal (4), Concentrations >2x water solubility (4), Organisms randomized (1), Exposure type appropriate (2), Hypothesis tests (3).

Daphnia magna

Study: Hamer MJ, Farrelly E, Hill IR. 1985b. PP321: 21 Day *Daphnia magna* life-cycle study. DPR report number 50907-089. ICI Plant Protection Division, Berkshire UK.

RelevanceReliabilityScore: 90 (No standard method)Score: 84.5Rating: RRating: R

Reference	Hamer et al. 1985b	D. magna
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth	< 24 h	
phase		
Source of organisms	Lab culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR at initiation, healthiest-	
	looking chosen at Day 6	
Test vessels randomized?	NR	
Test duration	21 d	
Data for multiple times?	Yes, 9d	
Effect 1	Mortality	
Control response 1	5d: 93%, 6-21 d: 90%	
Effect 2	Length of adults	
Control response 2	9 d: 4.75 mm, 21 d: 4.87 mm	
Effect 3	Total young produced	
Control response 3	3104	
Effect 4	Number of young/female/day	
Control response 4	7.28	
Effect 5	Number of female	
	reproductive days	
Control response 5	426 d	
Temperature	20 ± 1 °C	
Test type	Static renewal	renewed every 12 h

Reference	Hamer et al. 1985b	D. magna
Parameter	Value	Comment
Photoperiod/light intensity	16 L: 8 D, 800 lux	
Dilution water	Dechlorinated tap water	
рН	7.7-8.4	
Hardness	275 mg/L	
Alkalinity	245 mg/L	
Conductivity	665 μS/cm	
Dissolved Oxygen	> 7.9 mg/L (> 86% sat.)	
Feeding	Twice daily in <i>new</i> test solution, <i>Chlorella</i> and yeast.	Would have been better to feed right before changing new solution to avoid sorption to food and dietary exposure
Purity of test substance	99.6% radiochemical purity	Determined by TLC
Concentrations measured?	Yes	j
Measured is what % of nominal?	2.5 ng/L: t ₀ : 136%, 12 h: 150% All other conc: t ₀ : 85-98%, 12 h: 57-67%	
Chemical method documented?	Yes, LSC	TLC and HPLC to measure other aspects
Concentration of carrier (if any) in test solutions	0.005%	
Concentration 1 Nom/t ₀ Meas/12 h Meas (ng/L)	2.5/3.75/3.4	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
Concentration 2 Nom/ t ₀ Meas/12 h Meas (ng/L)	5/4.9/3.1	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
Concentration 3 Nom/ t ₀ Meas/12 h Meas (ng/L)	10/8.5/5.7	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
Concentration 4 Nom/ t ₀ Meas/12 h Meas (ng/L)	20/18.3/13.4	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
Concentration 5 Nom/ t ₀ Meas/12 h Meas (ng/L)	40/37.2/25	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
Control	Dil. water and Solvent	2 reps, 0-5 d: 50 org/rep, 6-21 d: 30 females/rep
NOEC	Total young: 4.9 ng/L	Method: 1-way

Reference	Hamer et al. 1985b	D. magna
Parameter	Value	Comment
	Female repro days: 8.5 ng/L	ANOVA
	Young/female/d: 3.75 ng/L	p: 0.05
	9 d length: 18.3 ng/L	MSD: NR
	21 d length: \geq 18.3 ng/L (not	
	enough surviving to assess at	
	40 ug/L)	
	5d mortality: not calculable	
	6-21 d mortality: not	
	calculable	
LOEC	Total young: 8.5 ng/L	Same as above
	Female repro days: 18.3 ng/L	
	Young/female/d: 4.9 ng/L	
	9 d length: 37.2 ng/L	
	21 d length: > 18.3 ng/L (not	
	enough surviving to assess at	
	40 ug/L)	
	5 d mortality: not calculable	
	6-21 d mortality: not	
	calculable	
MATC (GeoMean NOEC,LOEC)	Total young: 6.45 ng/L	
	Female repro days: 12.5 ng/L	
	Young/female/d: 4.3 ng/L	
	9 d length: 26.1 ng/L	
	21 d length: not calculable	
% control at NOEC	Total young: 73.7%	
	Female repro days: 97.8%	
	Young/female/d: 85.1%	
	9 d length: 91.6%	
	21 d length: 98.3%	
% of control LOEC	Total young: 67.7%	
	Female repro days: 81.9%	
	Young/female/d: 73.5%	
	9 d length: 86.1%	
	21 d length: not calculable	

- NOEC and LOEC determined from Table 5 based on statistical difference at p = 0.05.
- NOEC, LOEC, and MATC are based on measured concentrations at t₀.
- The NOEC/LOEC were not recorded for mortality at 5 or 21 d because no statistical calculations were done on the raw data.
- Some isomerization and hydrolysis did occur during the test as demonstrated by HPLC and TLC of the solutions.
- -After 5 d, the reps were reduced from 50 to 30 organisms by selecting only the healthiest looking females.
- There was no effect on the fertility of offspring transferred to untreated water for 13 d.

Reliability points taken off for:

<u>Documentation:</u> Statistical significance (2), Minimum significant difference (2), Point estimates (8)

Acceptability: Standard method (5), Organisms randomized (1), Appropriate feeding (3), Random design (2), Adequate replication (2), Point estimates (3).

Daphnia magna

Study: Machado MW. 2001a. XDE-225 and Lambda-cyhalothrin: Comparative toxicity to Daphnids (*Daphnia magna*) under static-renewal conditions. EPA MRID 45447220. Springborn Laboratories, Inc, Wareham, MA.

RelevanceReliabilityScore: 100Score: 91.5Rating: RRating: R

Reference	Machado 2001a	D. magna
Parameter	Value	Comment
Test method cited	US EPA 1996, OECD	
Phylum	Arthropoda	
Class	Branchiopoda	
Order	Cladocera	
Family	Daphniidae	
Genus	Daphnia	
Species	magna	
Family in North America?	Yes	
Age/size at start of test/growth	≤ 24 hr	
phase		
Source of organisms	Lab culture	Springborn labs
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	Yes	
Test duration	48 hr	
Data for multiple times?	Yes, 24 hr	
Effect 1	Immobilization	
Control response 1	0% at all times	
Temperature	21 ± 1 °C	
Test type	Static Renewal	Renewed at 24 hr
Photoperiod/light intensity	16 L: 8D, 50-80 footcandles	
Dilution water	Fortified well water	EPA hard water
pН	8.0-8.1	
Hardness	170 mg/L as CaCO ₃	
Alkalinity	120 mg/L as CaCO ₃	
Conductivity	500 μmhos/cm	
Dissolved Oxygen	8.4-8.9 mg/L (94-100% sat)	
Feeding	None during test	
Purity of test substance	99%	

Reference	Machado 2001a	D. magna
Parameter	Value	Comment
Concentrations measured?	Yes	
Measured is what % of nominal?	39.5-50%	
Chemical method documented?	Yes, GC/NCI-MS	
Concentration of carrier (if any) in	0.01%	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.013/0.0055	2 reps, 10 orgs/rep
Concentration 2 Nom/Meas (µg/L)	0.025/0.012	2 reps, 10 orgs/rep
Concentration 3 Nom/Meas (µg/L)	0.050/0.023	2 reps, 10 orgs/rep
Concentration 4 Nom/Meas (µg/L)	0.10/0.050	2 reps, 10 orgs/rep
Concentration 5 Nom/Meas (µg/L)	0.20/0.079	2 reps, 10 orgs/rep
Control	Solvent and dil. water	2 reps, 10 orgs/rep
EC ₅₀ (95% confidence interval)	24 h: >0.079 μg/L	Method: Probit
	48 h: 0.051 (0.034-0.10) μg/L	

EC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Adequate replication (2),

Hypothesis tests (3)

Gammarus pulex

Study: Hamer MJ, Farrelly E, Hill IR. 1985a. PP321: Toxicity to *Gammarus pulex*. ICI Plant Protection Division. DPR study number 50907-086.

RelevanceReliabilityScore: 100Score: 87.5Rating: RRating: R

Reference	Hamer et al. 1985a	G. pulex
Parameter	Value	Comment
Test method cited	ASTM 1980	
Phylum	Arthropoda	
Class	Malacotraca	
Order	Amphipoda	
Family	Gammaridae	
Genus	Gammarus	
Species	pulex	
Family in North America?	Yes	
Age/size at start of test/growth	5 mm, > 3 weeks old (exact	
phase	instar/age not given or	
	known)	
Source of organisms	River Wye, England	
Have organisms been exposed to contaminants?	Maybe	
Animals acclimated and disease-	Yes, for 3 weeks	
free?		
Animals randomized?	NR	
Test vessels randomized?	Yes	
Test duration	96 h	
Data for multiple times?	Yes, 24, 48, 72 h	
Effect 1	Immobility	
Control response 1	Tests 1 & 2: 0%	
Effect 2	Mortality	
Control response 2	Test 1: 5% at 72/96 h	
	Test 2: 0%	
Temperature	$15 \pm 0.5^{\circ}$ C	
Test type	Flow-through	
Photoperiod/light intensity	16 L: 8 D, 700 lux	
Dilution water	Dechlorinated tap water	
рН	8.1-8.4	
Hardness	250 mg/L	
Alkalinity	250 mg/L	
Conductivity	660 μS/cm	

Reference	Hamer et al. 1985a	G. pulex
Parameter	Value	Comment
Dissolved Oxygen	>9.3 mg/L (>91% sat.)	
Feeding	None during test	
Purity of test substance	99.2%	
Concentrations measured?	Yes	
Measured is what % of nominal?	Test 1: mean 72-86% (range	Meas. at 0, 24, 48,
	54-109%)	72, 96 h
	Test 2: mean 45-96% (range	,
	20-109%)	
Chemical method documented?	Yes, LSC	TLC to measure
		hydrolysis, HPLC
		to measure
		isomerization
Concentration of carrier (if any) in	None used	
test solutions		
Test 1 Concentration 1 Nom/Meas	Test 1: 65.5/54	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 39.5	org/rep
Test 1 Concentration 2 Nom/Meas	Test 1: 38.0/29.3	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 21.3	org/rep
Test 1 Concentration 3 Nom/Meas	Test 1: 20.5/14.6	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 10.0	org/rep
Test 1 Concentration 4 Nom/Meas	Test 1: 11.3/6.8	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 5.5	org/rep
Test 1 Concentration 5 Nom/Meas	Test 1: 5.3/3.9	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 3.4	org/rep
Test 1 Concentration 6 Nom/Meas	Test 1: 3.5/2.1	1 rep/test, 20
Test 2 Meas (ng/L)	Test 2: 2.0	org/rep
Test 1 Concentration 7 Nom/Meas	Test 1: 1.8/1.2	1 rep/test, 20
Test 2 Meas (ng/L) Test 1 Concentration 8 Nom/Meas	Test 2: 1.0 Test 1: 0.7/0.5	org/rep
		1 rep/test, 20 org/rep
Test 2 Meas (ng/L) Control	Test 2: 0.4 Dilution water	1 rep/test, 20
Control	Dilution water	org/rep
LC ₅₀ (95% confidence interval)	Test 1	Method: weighted
(ng/L)	24 h: 854 (133-infinity)	linear regression
(lig/L)	48 h: 55.4 (32.6-127)	inical regression
	72 h:26.9 (18.0-47.8)	
	96 h: 11.7 (8.2-17.6)	
	(0.2 17.0)	
	Test 2	
	24 h: 516 (0-infinity)	
	48 h: 95.0 (43.6-962)	
	72 h: 36.4 (24.3-74.3)	
	96 h: 13.8 (10.4-19.3)	
	, , ,	<u> </u>

Reference	Hamer et al. 1985a	G. pulex
Parameter	Value	Comment
	Mean of 2 tests	
	24 h: 665	
	48 h: 71.2	
	72 h: 31.3	
	96 h: 12.7	
EC ₅₀ (95% confidence interval)	Test 1	Method: weighted
(ng/L)	24 h: 8.9 (6.6-12.1)	linear regression
	48 h: 6.9 (3.4-14.9)	
	72 h: 6.1 (1.9-21.3)	
	96 h: 5.9 (2.1-18.2)	
	Test 2	
	24 h: 11.6 (5.8-36.4)	
	48 h: 9.1 (7.1-12.1)	
	72 h: 6.8 (5.2-9.0)	
	96 h: 5.9 (3.2-11.5)	
	Mean of 2 tests	
	24 h: 10.2	
	48 h: 8.0	
	72 h: 6.4	
	96 h: 5.9	

LC50 and EC50 calculated based on mean measured concentrations.

Some hydrolysis and isomerization did occur, but l-cyhalothrin always accounted for the majority of radioactivity.

Reliability points taken off for:

<u>Documentation:</u> Hypothesis tests (8)

<u>Acceptability:</u> Standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate age of organisms (3), Prior contamination (4), Organisms randomized (1), Adequate replication (2), Hypothesis tests (3).

Gammarus pulex

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	G. pulex
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Malacostraca	
Order	Amphipoda	
Family	Gammaridae	
Genus	Gammarus	
Species	pulex	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Pond	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	8.4-8.6	
Hardness	179 mg/L	
Alkalinity	150 mg/L	
Conductivity	NR	

Reference	Hamer <i>et al.</i> 1998	G. pulex
Parameter	Value	Comment
Dissolved Oxygen	8.2-9.1 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 99-122%; 48 h: 45-81%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	16/18/10	1 rep, 10 orgs/rep
(µg/L)		
Concentration 2 Nom/Meas t ₀ /48 h	31/31/25	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	62/76/37	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	125/131/59	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	250/247/112	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	500/534/266	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	14 (9.1-19) ng/L	Method: Iteratively
		re-weighted linear
		regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8) <u>Acceptability:</u> No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), No prior contamination (4), Organisms randomized (1), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Gasterosteus aculeatus

Study: Long KWJ, Shillabeer N. 1997a. Lambda-cyhalothrin: Acute toxicity to the three-spined stickleback (*Gasterosteus aculeatus*). DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Long & Shillabeer 1997a	G. aculeatus
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Chordata	
Class	Actinopterygii	
Order	Gasterosteiformes	
Family	Gasterosteidae	
Genus	Gasterosteus	
Species	aculeatus	Three-spined stickleback
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	-Mean weight and length at	
	end of test were 0.41 g and	
	34 mm	
	-> 2 weeks old	
Source of organisms	Lab culture	Blades Biological, Kent, UK
Have organisms been exposed to	No	
contaminants?	Yes	
Animals acclimated and disease-free?	Yes	
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	12 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
pН	7.03-7.31	
Hardness	46.0-47.3 mg/L	
Alkalinity	26.4 mg/L	

Reference	Long & Shillabeer 1997a	G. aculeatus
Parameter	Value	Comment
Conductivity	215-217 μS/cm	
Dissolved Oxygen	10.0-10.8 mg/L	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	68-138%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.10/0.068	1 rep, 23 org/rep
Concentration 2 Nom/Meas (µg/L)	0.20/0.16	1 rep, 23 org/rep
Concentration 3 Nom/Meas (µg/L)	0.40/0.68	1 rep, 23 org/rep
Concentration 4 Nom/Meas (µg/L)	0.80/0.79	1 rep, 23 org/rep
Concentration 5 Nom/Meas (µg/L)	1.6/1.5	1 rep, 23 org/rep
Concentration 6 Nom/Meas (µg/L)	3.2/2.5	1 rep, 23 org/rep
Control	Solvent and dilution water	1 rep, 23 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.73 (0.68-0.79)	Method: Binomial
$(\mu g/L)$	48 h: 0.44 (0.36-0.56)	(24 h), Moving
	72 h: 0.43 (0.35-0.54)	average angle (48,
	96 h: 0.40 (0.33-0.50)	72, 96 h)

 LC_{50} calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age at beginning (5), Photoperiod (3), Hypothesis tests (8) <u>Acceptability:</u> Measured concentrations w/in 20% of nominals (4), Appropriate age of organism (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Hyalella azteca

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 83.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	H. azteca
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Malacotraca	
Order	Amphipoda	
Family	Hyalellidae	
Genus	Hyalella	
Species	azteca	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Lab culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	7.7-8.6	
Hardness	179 mg/L	
Alkalinity	150 mg/L	
Conductivity	NR	

Reference	Hamer <i>et al.</i> 1998	H. azteca
Parameter	Value	Comment
Dissolved Oxygen	7.5-8.3 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 98-131%; 48 h: 56-64%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in test solutions	0.05%	
Concentration 1 Nom/Meas t ₀ /48 h (µg/L)	0.49/<10/<10	1 rep, 10 orgs/rep
Concentration 2 Nom/Meas t ₀ /48 h (ng/L)	1.0/<10/<10	1 rep, 10 orgs/rep
Concentration 3 Nom/Meas t ₀ /48 h (ng/L)	2.0/10/<10	1 rep, 10 orgs/rep
Concentration 4 Nom/Meas t ₀ /48 h (ng/L)	3.9/<10/<10	1 rep, 10 orgs/rep
Concentration 5 Nom/Meas t ₀ /48 h (ng/L)	7.8/10/<10	1 rep, 10 orgs/rep
Concentration 6 Nom/Meas t ₀ /48 h (ng/L)	16/16/10	1 rep, 10 orgs/rep
Concentration 7 Nom/Meas t ₀ /48 h (ng/L)	31/37/20	1 rep, 10 orgs/rep
Concentration 8 Nom/Meas t ₀ /48 h (ng/L)	62/61/35	1 rep, 10 orgs/rep
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	2.3 (1.0-7.8) ng/L	Method: Iteratively re-weighted linear regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Hyalella azteca

Study: Smith S, Lizotte RE. 2007. Influence of Selected Water Quality Characteristics on the Toxicity of λ -cyhalothrin and γ -cyhalothrin to *Hyalella azteca*. Bull Environ Contam Toxicol, 79:548-551.

RelevanceReliabilityScore: L (Low chemical purity)Score: RRating: 85Rating: 76.5

Reference	Smith & Lizotte 2007	H. azteca
Parameter	Value	Comment
Test method cited	US EPA 1994	
Phylum	Arthropoda	
Class	Malacotraca	
Order	Amphipoda	
Family	Hyalellidae	
Genus	Hyalella	
Species	azteca	
Family in North America?	Yes	
Age/size at start of test/growth	1-2 weeks old	
phase	$< 600 \ \mu m, \ge 425 \ \mu m$	
Source of organisms	Lab culture	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	99 ± 1%	
Temperature	23± 1°C	
Test type	Static	
Photoperiod/light intensity	16 L: 8 D	
Dilution water	Fortified, unfiltered pond	
	waters	
рН	8.0	
Hardness	93 mg/L	
Alkalinity	55 mg/L	
Conductivity	360 μmhos/cm	
Dissolved Oxygen	7.7 mg/L	
Feeding	None during test	

Reference	Smith & Lizotte 2007	H. azteca
Parameter	Value	Comment
Purity of test substance	22.8% active ingredient in	
	formulation stock	
Concentrations measured?	Stock solutions measured	
Measured is what % of nominal?	31-75%	
Chemical method documented?	Yes, GC	
Concentration of carrier (if any) in	NR, but formulation was	
test solutions	used so several other	
	chemicals were present	
Concentration 1 Nom/Meas (µg/L)	0.0009	6 reps, 5 orgs/rep
Concentration 2 Nom/Meas (µg/L)	NR	6 reps, 5 orgs/rep
Concentration 3 Nom/Meas (µg/L)	NR	6 reps, 5 orgs/rep
Concentration 4 Nom/Meas (µg/L)	NR	6 reps, 5 orgs/rep
Concentration 5 Nom/Meas (µg/L)	0.6	6 reps, 5 orgs/rep
Control	Dilution water	6 reps, 5 orgs/rep
48 h EC ₅₀ (95% confidence	1: 2.8 (2.0-3.8) ng/L	Method: Probit (if
interval) – for 12 different waters	2: 1.7 (1.3-2.2) ng/L	linear), Trimmed
tested	3: 2.4 (1.8-3.1) ng/L	Spearman-Karber
	4: 10.4 (8.3-13.6) ng/L	(if non-linear)
	5: 1.5 (1.1-1.9) ng/L	
	6: 7.4 (5.9-9.2) ng/L	
	7: 3.9 (3.0-4.9) ng/L	
	8: 1.4 (1.1-1.8) ng/L	
	9: 3.6 (2.8-4.5) ng/L	
	10: 2.2 (1.7-2.8) ng/L	
	11: 11.1 (8.7-14.3) ng/L	
	12: 15.7 (12.5-19.7) ng/L	

EC50 values calculated based on nominal concentrations.

Mean water quality characteristics of 12 waters (Std dev)

Water body	Turbidity	TSS (mg/L)	Chlorophyll a	DOC (mg/L)
-	(NTU)		(ug/L)	
1	7.0 (0.3)	10 (6)	3.8 (2.5)	10.6 (2.4)
2	3.1 (0.2)	2(1)	2.9 (1.7)	1.4 (0.2)
3	0.7 (0.2)	4 (3)	5.3 (2.5)	1.0 (0.2)
4	6.6 (0.5)	13 (4)	35.9 (3.7)	16.7 (5.8)
5	1.2 (0.5)	4 (2)	1.7 (1.7)	1.9 (0.2)
6	16.0 (0.5)	15 (5)	76.1 (2.3)	11.4 (3.7)
7	3.5 (0.2)	5 (3)	3.0 (1.8)	2.7 (0.2)
8	1.9 (0.2)	1(1)	1.7 (1.2)	2.8 (0.4)
9	2.0 (0.2)	6 (3)	1.2 (1.3)	5.3 (1.0)

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10	1.4 (0.1)	2(1)	5.6 (3.2)	1.7 (0.3)
11	19.9 (0.6)	14 (7)	36.2 (2.1)	11.4 (1.8)
12	67.2 (2.0)	79 (5)	102.0 (5.8)	32.9 (10.2)

Linear regression relationships between water quality parameters and toxicity:

Turbidity(x, NTU): EC_{50} =0.216x + 3.04, R^2 =0.712, F = 24.7, p = 0.0006 TSS (x, mg/L): EC_{50} = 0.179x + 3.15, R^2 = 0.644, F = 18.1, p = 0.0017 DOC (x, mg/L): EC_{50} = 0.546x + 1.07, R^2 = 0.847, F = 55.3, p < 0.0001 Chlorophyll a (x, µg/L): EC_{50} = 0.123x + 2.61, R^2 = 0.742, F = 28.7, p = 0.0003

Interaction of increased DOC and phytoplankton (as chl a) decreases toxicity of l-cyhalothrin to H. azteca by more than 10-fold.

Reliability points taken off for:

Documentation: Hypothesis tests (8)

Acceptability: Appropriate control (6), Chemical purity (10), Measured concentrations w/in 20% of nominal (4), Carrier solvent (4), Organisms randomized (1), Exposure type (2), Dilution water (2), Number of concentrations (3), Random design (2), Dilution factor (2), Hypothesis tests (3)

Hydracarina

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	Hydracarina
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Arachnida	
Order	Trombidformes	
Suborder	Hydracarina*	
Genus	NR	
Species	NR	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
	research station	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	8.0-8.7	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	Hydracarina
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.4-9.0 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 110-119%; 48 h: 60-69%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	31/37/20	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	62/74/37	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	125/147/86	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	250/288/168	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	500/550/313	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	1000/1139/646	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	47 (33-62) ng/L	Method: Iteratively
		re-weighted linear
		regression

* The binomial name was not available, as these organisms were collected from the field and species identification is very difficult and not well-described in the literature. There were no other data available for similar organisms, so there is no question about combining values for a species mean value and the variability (as shown by the 95% confidence interval) is low, so it is reasonable to believe that the test organisms were all one species.

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Conductivity (2), Hypothesis tests (8) <u>Acceptability:</u> No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Prior contaminant exposure (4), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Ictalurus punctatus

Study: Long KWJ, Shillabeer N. 1997b. Lambda-cyhalothrin: Acute toxicity to channel catfish (*Ictalurus punctatus*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Long & Shillabeer 1997b	I. punctatus
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Siluriformes	
Family	Ictaluridae	
Genus	Ictalurus	
Species	punctatus	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	-Mean weight and length at	
	end of test were 1.57 g and	
	48 mm	
	- > 25 d old	
Source of organisms	Lab culture	Aquatic Research
		Organisms, Hampton, NH, USA
Have organisms been exposed to	No	,,
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	17 ± 1°C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
pH	7.24-7.70	
Hardness	43.0-47.7 mg/L	
Alkalinity	26.7 mg/L	

Reference	Long & Shillabeer 1997b	I. punctatus
Parameter	Value	Comment
Conductivity	201-211 μS/cm	
Dissolved Oxygen	8.8-10.0 mg/L	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	25-57%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.080/0.020	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.16/0.058	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.32/0.090	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.64/0.25	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	1.28/0.73	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	2.56/1.0	1 rep, 20 org/rep
Control	Solvent and dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.82 (0.67-1.1)	Method: Binomial
$(\mu g/L)$	48 h: 0.43 (0.25-0.73)	(48 h), Moving
	72 h: 0.18 (0.15-0.23)	average angle (24,
	96 h: 0.16 (0.13-0.20)	72, 96 h)

 LC_{50} calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age at beginning (5), Photoperiod (3), Hypothesis tests (8) <u>Acceptability:</u> Measured concentrations w/in 20% of nominals (4), Appropriate age of organism (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Ischnura elegans

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 81.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	I. elegans
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Class	Insecta	
Order	Odonata	
Family	Coenagrionidae	
Genus	Ischnura	
Species	elegans	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Experimental ponds at	
-	research stations	
Have organisms been exposed to	Maybe	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	8.2-8.7	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	I. elegans
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.6-9.0 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 87-116%; 48 h: 50-82%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in test solutions	0.05%	
Concentration 1 Nom/Meas t ₀ /48 h (µg/L)	31/27/22	1 rep, 10 orgs/rep
Concentration 2 Nom/Meas t ₀ /48 h (ng/L)	62/72/51	1 rep, 10 orgs/rep
Concentration 3 Nom/Meas t ₀ /48 h (ng/L)	125/125/63	1 rep, 10 orgs/rep
Concentration 4 Nom/Meas t ₀ /48 h (ng/L)	250/229/129	1 rep, 10 orgs/rep
Concentration 5 Nom/Meas t ₀ /48 h (ng/L)	500/436/313	1 rep, 10 orgs/rep
Concentration 6 Nom/Meas t ₀ /48 h (ng/L)	1000/1031/701	1 rep, 10 orgs/rep
Concentration 7 Nom/Meas t ₀ /48 h (ng/L)	2000/1857/1137	1 rep, 10 orgs/rep
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	38 (23-93) ng/L	Method: Iteratively re-weighted linear regression

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

Documentation: Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Prior contamination (4),

Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Lepomis macrochirus

Study: Hill RW. 1984b. PP321: Determination of acute toxicity to bluegill sunfish (*Lepomis macrochirus*). ICI Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 90 (No standard method)Score: 83Rating: RRating: R

Reference	Hill 1984b	L. macrochirus
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Chordata	
Class	Actinopterygii	
Order	Perciformes	
Family	Centrarchidae	
Genus	Lepomis	
Species	macrochirus	
Family in North America?	Yes	
Age/size at start of test/growth	Mean wt.: 1.51 g	
phase	Mean length: 38.2 mm	
Source of organisms	Commercial – Sea	
-	Plantations Inc. Salem, MA	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 96 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	22 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Freshwater reservoir	Not more specific
pH	7.4-8.6	
Hardness	68.2 mg/L	
Alkalinity	28.4mg/L	
Conductivity	137 μS/cm	
Dissolved Oxygen	7.0-8.4 mg/L	
Feeding	NR	
Purity of test substance	98%	

Reference	Hill 1984b	L. macrochirus
Parameter	Value	Comment
Concentrations measured?	Yes	
Measured is what % of nominal?	50-65%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	9 mg/L acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	1.8/1.17	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	1.0/0.65	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.56/0.31	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.32/0.16	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	0.18/0.10	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	0.10/0.06	1 rep, 20 org/rep
Control	Solvent and dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.45 (0.38-0.52)	Method: Probit
$(\mu g/L)$	48 h: 0.28 (0.23-0.32)	
	72 h: 0.28 (0.23-0.32)	
	96 h: 0.21 (0.18-0.25)	

 LC_{50} calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Dilution water source (3), Photoperiod (3), Hypothesis tests (8), <u>Acceptability:</u> Standard method (5), Measured concentrations w/in 20% nominal (4), Organisms randomized (1), Dilution water (2), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Lepomis macrochirus

Study: Marino TA, Rick DL. 2001. XR-225 and lambda-cyhalothrin: An acute toxicity comparison study with the Bluegill sunfish, *Lepomis macrochirus* Rafinesque. EPA MRID 45447216.

RelevanceReliabilityScore: 100Score: 90Rating: RRating: R

Reference	Marino & Rick 2001	L. macrochirus
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Actinopterygii	
Order	Perciformes	
Family	Centrarchidae	
Genus	Lepomis	
Species	macrochirus	Rafinesque
Family in North America?	Yes	
Age/size at start of test/growth	Juvenile	
phase		
Source of organisms	Laboratory culture	Northeaster
		Aquatics,
		Rhinebeck, NY
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	No	
Test duration	96 hr	
Data for multiple times?	Yes – 24 hr, 48 hr, 72 hr	
Effect 1	Mortality	
Control response 1	0% at all times	
Effect 2	<10% mortality	
Control response 2	0% mortality at all times	
Effect 3	Behavioral effects	
Control response 3	0% at all times	
Temperature	21.9 ± 0.3 °C	
Test type	Flow-through	
Photoperiod/light intensity	16 L: 8 D	
Dilution water	Filtered Lake Huron water	
pН	7.0 ± 0.1	

Reference	Marino & Rick 2001	L. macrochirus
Parameter	Value	Comment
Hardness	55 mg/L as CaCO ₃	
Alkalinity	30 mg/L as CaCO ₃	
Conductivity	71.1 μ mho/cm	
Dissolved Oxygen	$8.8 \pm 0.6 \text{ mg/L}$	≥87% saturation
Feeding	None during test	
Purity of test substance	99%	
Concentrations measured?	Yes	
Measured is what % of nominal?	44-58%	
Chemical method documented?	Yes	
Concentration of carrier (if any) in	0.8 mL/L (0.08%)	
test solutions		
Concentration 1 Nom/Meas (ng/L)	27.2/13.7	2 reps, 5 fish/rep
Concentration 2 Nom/Meas (ng/L)	45.4/20.8	2 reps, 5 fish/rep
Concentration 3 Nom/Meas (ng/L)	75.6/34.0	2 reps, 5 fish/rep
Concentration 4 Nom/Meas (ng/L)	126/65.4	2 reps, 5 fish/rep
Concentration 5 Nom/Meas (ng/L)	210/104	2 reps, 5 fish/rep
Concentration 6 Nom/Meas (ng/L)	350/203	2 reps, 5 fish/rep
Control	< detection limit (10 ng/L)	2 reps, 5 fish/rep
LC ₅₀	24 h: 224 (152-1742) ng/L	Method: Probit
	48 h: 124 (94.4-163) ng/L	and/or Trimmed
	72 h: 118 (94.4-155) ng/L	Spearman-Karber
	96 h: 106 (85.5-140) ng/L	

Point estimates based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% nominal (4), Carrier solvent (4), Random design (2), Adequate replication (2), Hypothesis tests (3)

Leuciscus idus

Study: Kent SJ, Shillabeer N. 1997a. Lambda-cyhalothrin: Acute toxicity to golden orfe (*Leuciscus idus*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Kent & Shillabeer 1997a	L. idus
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Cypriniformes	
Family	Cyprinidae	
Genus	Leuciscus	
Species	idus	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	>17 d old	
	> mean control weight	
	and length 2.15 g and	
	53 mm at end of test.	
Source of organisms	Lab culture	London Aquatic
		Co., UK
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	17 d acclimation in
free?		facility
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	12 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
рН	7.04-7.39	
Hardness	43.3-46.3 mg/L	
Alkalinity	25 mg/L	
Conductivity	212-218 μS/cm	

Reference	Kent & Shillabeer 1997a	L. idus
Parameter	Value	Comment
Dissolved Oxygen	9.2-10.4 mg/L, > 87% sat	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	43-58%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.030/0.017	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.060/0.026	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.12/0.056	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.24/0.11	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	0.48/0.28	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	0.96/0.48	1 rep, 20 org/rep
Control	Solvent and Dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.18 (0.11-0.28)	Method: Binomial
$(\mu g/L)$	48 h: 0.078 (0.056-0.11)	
	72 h: 0.078 (0.056-0.11)	
	96 h: 0.078 (0.056-0.11)	

LC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Photoperiod (3), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Macrobrachium nippoensis

Study: Wang W, Cai DJ, Shan ZJ, Chen WL, Poletika N, Gao XW. 2007. Comparison of the acute toxicity for gamma-cyhalothrin and lambda-cyhalothrin to zebra fish and shrimp. Regulatory Toxicology and Pharmacology, 47: 184-188.

RelevanceReliabilityScore: 75 (No standard method, low chemical purity)Score: 63Rating: LRating: L

Reference	Wang et al. 2007	M. nippoensis
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Arthropoda	
Class	Malacostraca	
Order	Decapoda	
Family	Palaemonoidea	
Genus	Macrobrachium	
Species	nippoensis	
Family in North America?	Yes	
Age/size at start of test/growth phase	90 d old, 5.0 g, 4.5 cm	
Source of organisms	Lab culture	Nanjing Institute of Environmental Sciences
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes, 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	16 ± 2 °C	
Test type	Static renewal, renewed	
	every 24 h	
Photoperiod/light intensity	NR	
Dilution water	NR	
рН	7.1	
Hardness	6.8-8.0 °HG	
Alkalinity	NR	
Conductivity	NR	

Reference	Wang et al. 2007	M. nippoensis
Parameter	Value	Comment
Dissolved Oxygen	NR	
Feeding	None during test	
Purity of test substance	Kung Fu 25 EW formulation	
Concentrations measured?	Yes, but NR	
Measured is what % of nominal?	NR	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	None used	
test solutions		
Concentration 1 Nom/Meas (a.i.	0.02	1 rep, 10 orgs/rep
μg/L)		
Concentration 2 Nom/Meas (a.i.	0.03	1 rep, 10 orgs/rep
μg/L)		
Concentration 3 Nom/Meas (a.i.	0.05	1 rep, 10 orgs/rep
µg/L)		
Concentration 4 Nom/Meas (a.i.	0.10	1 rep, 10 orgs/rep
µg/L)		
Concentration 5 Nom/Meas (a.i.	0.20	1 rep, 10 orgs/rep
μg/L)		
Control	Dilution water	1 rep, 10 orgs/rep
LC ₅₀ (95% confidence interval)	24 h: 0.05 (0.04-0.07)	Method: NR
(µg/L)	48 h: 0.05 (0.04-0.06)	
	72 h: 0.04 (0.03-0.06)	
	96 h: 0.04 (0.03-0.05)	

LC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Chemical purity (5), Measured concentrations (3), Dilution water (3), Alkalinity (2), Dissolved oxygen (4), Conductivity (2), Photoperiod (3), Statistical methods (5), Hypothesis tests (8),

Acceptability: Standard method (5), Chemical purity (10), Measured concentrations w/in 20% nominal (4), Organisms randomized (1), Alkalinity (2), Dissolved oxygen (6), Conductivity (1), Photoperiod (2), Random design (2), Adequate replication (2), Statistical method (2), Hypothesis tests (3).

Mysidopsis bahia

Study: Thompson RS. 1985. PP321: Determination of acute toxicity to mysid shrimp (*Mysidopsis bahia*). DPR study number 50907-0087, 160359.

RelevanceReliabilityScore: 77.5 (Saltwater, low solv. control response)Score: 79Rating: LRating: R

Reference	Thompson 1985	M. bahia
Parameter	Value	Comment
Test method cited	EPA GLP	
Phylum	Arthropoda	
Class	Malacostraca	
Order	Mysida	
Family	Vespoidea	
Genus	Mysidopsis	
Species	bahia	
Family in North America?	Yes	
Age/size at start of test/growth	< 48 h	
phase		
Source of organisms	Commercial supplier, Sea	
_	Plantations, Inc.	
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	24, 48, 72 h	
Effect 1	Mortality	
Control response 1	Dil: 0%, Solv: 15%	
Temperature	25 ± 1 °C	
Test type	FT	
Photoperiod/light intensity	14 L: 10 D	
Dilution water	Natural seawater diluted with	Seawater from Tor
	freshwater	Bay, UK
рН	8.12-8.22	
Hardness	NR	
Alkalinity	NR	
Salinity	20 o/oo	
Dissolved Oxygen	6.6-7.4 mg/L	
Feeding	Yes, twice daily	

Reference	Thompson 1985	M. bahia
Parameter	Value	Comment
Purity of test substance	97%	
Concentrations measured?	Yes	
Measured is what % of nominal?	29-81%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.022 mL/L	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.0032/0.0026	1 rep, 20 orgs/rep
Concentration 2 Nom/Meas (µg/L)	0.0056/0.0026	1 rep, 20 orgs/rep
Concentration 3 Nom/Meas (µg/L)	0.010/0.0059	1 rep, 20 orgs/rep
Concentration 4 Nom/Meas (µg/L)	0.018/0.0052	1 rep, 20 orgs/rep
Concentration 5 Nom/Meas (µg/L)	0.032/0.011	1 rep, 20 orgs/rep
Concentration 6 Nom/Meas (µg/L)	0.056/0.0166	1 rep, 20 orgs/rep
Control	Dilution water and solvent	1 rep, 20 orgs/rep
LC ₅₀ (95% confidence interval)	24 h: >0.017	Method: Probit
$(\mu g/L)$	48 h: 0.0075 (0.0061-0.0096)	
	72 h: 0.0049 (0.0041-0.0058)	
	96 h: 0.0041 (0.0034-0.0049)	

LC₅₀ calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hardness (2), Alkalinity (2), Conductivity (2), Hypothesis tests (8). <u>Acceptability:</u> Control response not acceptable (9), Measured concentrations w/in 20% nominal (4), Organism feeding (3), Hardness (2), Alkalinity (2), Conductivity (1), Random design (2), Adequate replication (2), Hypothesis tests (3).

Mysidopsis bahia

Study: Thompson . 1987. PP321 (Lambda-cyhalothrin): Determination of chronic toxicity to mysid shrimps (*Mysidopsis bahia*). DPR study 50907-089.

RelevanceReliabilityScore: 77.5 (Saltwater, Control Response)Score: 79Rating: LRating: R

Reference	Thompson 1987	M. bahia
Parameter	Value	Comment
Test method cited	ASTM 1986	
Phylum	Arthropoda	
Class	Malacotraca	
Order	Mydida	
Family	Vespoidea	
Genus	Mysidopsis	
Species	bahia	
Family in North America?	Yes	
Age/size at start of test/growth phase	< 24 g	
Source of organisms	Continuous lab culture at	
C	testing facility	
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	28 d	
Data for multiple times?	yes, 14, 12 d	
Effect 1	Mortality	
Control response 1	Dil water: 22.5%, Solvent: 12.5%	
Effect 2	Dry weight	
Control response 2	Female: 0.96 mg, Male: 0.81 mg	
Effect 3	Reproduction	
Control response 3	0.9 young/available female/day	
Temperature	$25 \pm 1^{\circ}$ C	
Test type	Flow-through	
Photoperiod/light intensity	14 L: 10 D	
Dilution water	Mixture of filtered seawater	

Reference	Thompson 1987	M. bahia
Parameter	Value	Comment
	and freshwater	
pН	7.90-8.20	
Hardness	NR	
Alkalinity	NR	
Salinity	20 o/oo	
Dissolved Oxygen	6.35-7.55 mg/L	
Feeding	Daily	
Purity of test substance	98.5%	
Concentrations measured?	Yes	
Measured is what % of nominal?	48-74%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.00037% triethylene glycol	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.00033/0.00022	2 reps, 20 orgs/rep
Concentration 2 Nom/Meas (µg/L)	0.00065/0.00046	2 reps, 20 orgs/rep
Concentration 3 Nom/Meas (µg/L)	0.0013/0.00070	2 reps, 20 orgs/rep
Concentration 4 Nom/Meas (µg/L)	0.0025/0.0017	2 reps, 20 orgs/rep
Concentration 5 Nom/Meas (µg/L)	0.005/0.0037	2 reps, 20 orgs/rep
Concentration 6 Nom/Meas (µg/L)	0.010/0.0048	2 reps, 20 orgs/rep
Control	Solvent and dilution water	2 reps, 20 orgs/rep
NOEC (µg/L)	Reproduction, mortality, dry	Method: Dunnett's
	weight: 0.00022	test, Student t-test
		p: 0.05
		MSD: NR
LOEC (µg/L)	Reproduction: 0.00046	Same as above
MATC (GeoMean NOEC,LOEC)	Reproduction: 0.00032 µg/L	

NOEC/LOEC calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hardness (2), Alkalinity (2), Conductivity (2), Minimum significant difference (2), Point estimates (8)

Acceptability: Control response (9), Measured concentrations w/in 20% of nominal (4), Hardness (2), Alkalinity (2), Conductivity (1), Random design (2), Adequate replication (2), Minimum significant difference (1), Point estimates (3)

Oncorhynchus mykiss (formerly Salmo gairdneri)

Study: Hill RW. 1984a. PP321: Determination of acute toxicity to rainbow trout (*Salmo gairdneri*). ICI Agrochemicals. DPR 50907-008.

RelevanceReliabilityScore: 90 (No standard method)Score: 81Rating: RRating: R

Reference	Hill 1984a	O. mykiss
Parameter	Value	Comment
Test method cited	None cited	
Phylum	Chordata	
Class	Osteichthyes	
Order	Salmoniformes	
Family	Salmonidae	
Genus	Oncorhynchus	
Species	mykiss	
Family in North America?	Yes	
Age/size at start of test/growth	Mean weight: 0.83 g	
phase	Mean length: 38.3 mm	
Source of organisms	Continuous culture at testing facility	
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	12 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Freshwater reservoir	Not more specific
pН	7.7-7.9	
Hardness	72.4 mg/L	
Alkalinity	29.6 mg/L	
Conductivity	165 μS/cm	
Dissolved Oxygen	10.2-11.2 mg/L	
Feeding	NR	

Reference	Hill 1984a	O. mykiss
Parameter	Value	Comment
Purity of test substance	98%	
Concentrations measured?	Yes	
Measured is what % of nominal?	54-70%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	9 mg/L acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	1.0/0.63	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.56/0.37	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.32/0.21	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.18/0.11	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	0.10/0.07	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	0.056/0.03	1 rep, 20 org/rep
Control	Solvent and dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.52 (0.46-0.60)	Method: Probit
$(\mu g/L)$	48 h: 0.40 (0.35-0.45)	
	72 h: 0.27 (0.09-0.80)	
	96 h: 0.24 (0.08-0.70)	

LC₅₀ calculated based on mean measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Dilution water source (3), Photoperiod (3), Hypothesis tests (8), <u>Acceptability:</u> No standard method (5), Measured concentrations w/in 20% nominal (4), Organisms randomized (1), Feeding (3), Dilution water (2), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (8).

Oncorhynchus mykiss

Study: Machado MW. 2001b. XDE-225 and Lambda-cyhalothrin: Comparative toxicity to Rainbow Trout (*Oncorhynchus mykiss*) under flow-through conditions.

RelevanceReliabilityScore: 100Score: 90.5Rating: RRating: R

Reference	Machado 2001b	O. mykiss
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Salmoniformes	
Family	Salmonidae	
Genus	Oncorhynchus	
Species	mykiss	
Family in North America?	Yes	
Age/size at start of test/growth	39 mm, 0.52 g	
phase		
Source of organisms	Lab culture	Trout Lodge,
		Sumner, WA
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	
free?		
Animals randomized?	Yes	
Test vessels randomized?	No	
Test duration	96 hr	
Data for multiple times?	Yes – 24 h, 48 hr, 72 hr	
Effect 1	Mortality	
Control response 1	0% at all times	
Temperature	12 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	16 L: 8 D, 40-80 footcandles	
Dilution water	Well water	
рН	7.3	
Hardness	42-44 mg/L as CaCO ₃	
Alkalinity	34-35 mg/L as CaCO ₃	
Conductivity	140-150 μmhos/cm	
Dissolved Oxygen	8.6-10.0 mg/L (80-93% sat)	
Feeding	None during test	
Purity of test substance	99%	

Reference	Machado 2001b	O. mykiss
Parameter	Value	Comment
Concentrations measured?	Yes	
Measured is what % of nominal?	100-133%	
Chemical method documented?	Yes, GC/MS	
Concentration of carrier (if any) in	0.096 mL/L (0.0096%)	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.043/0.051	2 reps, 10 fish/rep
Concentration 2 Nom/Meas (µg/L)	0.071/0.078	2 reps, 10 fish/rep
Concentration 3 Nom/Meas (µg/L)	0.12/0.16	2 reps, 10 fish/rep
Concentration 4 Nom/Meas (µg/L)	0.20/0.20	2 reps, 10 fish/rep
Concentration 5 Nom/Meas (µg/L)	0.33/0.38	2 reps, 10 fish/rep
Control	0	2 reps, 10 fish/rep
LC ₅₀ (95% confidence interval)	24 h: >0.38 ug/L	Method: nonlinear
	48 h: 0.29 (0.25-0.33) ug/L	interpolation
	72 h: 0.22 (0.20-0.38) ug/L	
	96 h: 0.19 (0.16-0.20) ug/L	

LC50 calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Hypothesis tests (8)

<u>Acceptability:</u> Measured concentrations w/in 20% of nominal (4), Random design (2),

Adequate replication (2), Hypothesis tests (3)

Oncorhynchus mykiss (formerly Salmo gairdneri)

Study: Tapp JF, Sankey SA, Caunter JE, Harland BJ. 1989. Lambda-cyhalothrin: Determination of acute toxicity to rainbow trout (*Salmo gairdneri*). ICI Agrochemicals. DPR study 50907-085.

RelevanceReliabilityScore: 100Score: 86.5Rating: RRating: R

Reference	Tapp <i>et al.</i> 1989	O. mykiss
Parameter	Value	Comment
Test method cited	US EPA	
Phylum	Chordata	
Class	Osteichthyes	
Order	Salmoniformes	
Family	Salmonidae	
Genus	Oncorhynchus	Formerly Salmo
Species	mykiss	gairdneri
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	-mean weight and length of 1.12 g, 43 mm at end of test -> 19 d old	
Source of organisms	Commercial lab	Zeals Fish Farm, UK
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes, 19 d acclimatization	
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	12 ± 1°C	
Test type	Flow-through	
Photoperiod/light intensity	16 L: 8 D	
Dilution water	Dechlorinated tap water	
pН	7.4-7.8	
Hardness	63.0-65.7 mg/L	
Alkalinity	37.6 mg/L	

Reference	Tapp <i>et al.</i> 1989	O. mykiss
Parameter	Value	Comment
Conductivity	207-212 μS/cm	
Dissolved Oxygen	8.2-9.8 mg/L, >78 % sat	
Feeding	None during test	
Purity of test substance	81.5%	
Concentrations measured?	Yes	
Measured is what % of nominal?	59-82%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.0005% acetone	
test solutions		
Concentration 1 Nom/Meas (µg/L)	1.0/0.72	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.56/0.33	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.32/0.20	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.18/0.12	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	0.1/0.076	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	0.056/0.046	1 rep, 20 org/rep
Control	Solvent and dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: > 0.72	Method: Moving
$(\mu g/L)$	48 h: 0.57 (0.50-0.66)	average
	72 h: 0.49 (0.43-0.58)	
	96 h: 0.44 (0.38-0.51)	

 LC_{50} calculated based on mean measured concentrations.

Reliability points taken off for:

Documentation: Initial organism age (5), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Random design (2), Adequate replication (2), Hypothesis tests (3).

Oryzias latipes

Study: Kent SJ, Shillabeer N. 1997d. Lambda-cyhalothrin: Acute toxicity to Japanese rice fish (*Oryzias latipes*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 85 (not from N. America)Score: 84Rating: LRating: R

Reference	Kent & Shillabeer 1997d	O. latipes
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Actinopterygii	
Order	Beloniformes	
Family	Adrianichthyidae	
Genus	Oryzias	
Species	latipes	
Family in North America?	No	
Age/size at start of test/growth	NR	
phase	> >17 d old	
	mean control weight	
	and length 0.22 g and	
	25 mm at end of test.	
Source of organisms	Lab culture	Aquatic Research
		Organisms,
		Hampton, NH, USA
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-	Yes	17 d acclimation in
free?		facility
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	Dil water: 20%	
	Solvent: 0%	
Temperature	$25 \pm 1^{\circ}\text{C}$	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
pН	7.63-7.83	

Reference	Kent & Shillabeer 1997d	O. latipes
Parameter	Value	Comment
Hardness	40.0-48.7 mg/L	
Alkalinity	27.4 mg/L	
Conductivity	197-215 μS/cm	
Dissolved Oxygen	7.6-8.8 mg/L	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	16-58%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.30/0.047	1 rep, 10 org/rep
Concentration 2 Nom/Meas (µg/L)	0.60/0.24	1 rep, 10 org/rep
Concentration 3 Nom/Meas (µg/L)	1.2/0.25	1 rep, 10 org/rep
Concentration 4 Nom/Meas (µg/L)	2.4/0.95	1 rep, 10 org/rep
Concentration 5 Nom/Meas (µg/L)	4.8/2.5	1 rep, 10 org/rep
Concentration 6 Nom/Meas (µg/L)	9.6/5.6	1 rep, 10 org/rep
Control	Solvent and Dilution water	1 rep, 10 org/rep
LC ₅₀ (95% confidence interval)	24 h: 2.1 (1.5-3.3)	Method: Moving
$(\mu g/L)$	48 h: 1.5 (1.0-2.6)	average angle
	72 h: 1.4 (0.93-2.3)	
	96 h: 1.4 (0.93-2.3)	

LC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Photoperiod (3), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Ostracoda

Study: Hamer MJ, Ashwell JA, Gentle WE. 1998. Lambda-cyhalothrin Acute Toxicity to Aquatic Arthropods. ZENECA Agrochemicals, Jealott's Hill Research Station Bracknell, Berkshire, UK. DPR study number 50907-093.

RelevanceReliabilityScore: 100Score: 83.5Rating: RRating: R

Reference	Hamer <i>et al.</i> 1998	Ostracoda
Parameter	Value	Comment
Test method cited	USEPA, OECD	
Phylum	Arthropoda	
Subphylum	Crustacea	
Class	Ostracoda*	
Family	NR	
Genus	NR	
Species	NR	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase		
Source of organisms	Lab culture	From H. azteca
		culture
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	NR	
free?		
Animals randomized?	NR	
Test vessels randomized?	NR	
Test duration	48 h	
Data for multiple times?	No	
Effect 1	Immobility	
Control response 1	0%	
Effect 2	Mortality	
Control response 2	0%	
Temperature	20 ± 2 °C	
Test type	Static	
Photoperiod/light intensity	16 L:8 D, 700-1000 lux	
Dilution water	Mixture of dechlorinated and	
	RO filtered tap water	
рН	7.8-8.8	
Hardness	179 mg/L	
Alkalinity	150 mg/L	

Reference	Hamer <i>et al.</i> 1998	Ostracoda
Parameter	Value	Comment
Conductivity	NR	
Dissolved Oxygen	8.4-8.7 mg/L	
Feeding	None during test	
Purity of test substance	≥ 88% radiochemical purity	
Concentrations measured?	Yes	
Measured is what % of nominal?	t ₀ : 81-104%; 48 h: 37-43%	
Chemical method documented?	Yes, LSC	
Concentration of carrier (if any) in	0.05%	
test solutions		
Concentration 1 Nom/Meas t ₀ /48 h	250/202/100	1 rep, 10 orgs/rep
$(\mu g/L)$		
Concentration 2 Nom/Meas t ₀ /48 h	500/434/213	1 rep, 10 orgs/rep
(ng/L)		
Concentration 3 Nom/Meas t ₀ /48 h	1000/900/405	1 rep, 10 orgs/rep
(ng/L)		
Concentration 4 Nom/Meas t ₀ /48 h	2000/1988/838	1 rep, 10 orgs/rep
(ng/L)		
Concentration 5 Nom/Meas t ₀ /48 h	4000/4155/1722	1 rep, 10 orgs/rep
(ng/L)		
Concentration 6 Nom/Meas t ₀ /48 h	8000/8078/2988	1 rep, 10 orgs/rep
(ng/L)		
Control (ng/L)	Dil. water and solvent: <10	1 rep, 10 orgs/rep
EC ₅₀ (95% confidence interval)	3300 (2100-6600) ng/L	Method: Iteratively
		re-weighted linear
		regression

^{*} The binomial name was not available, as these organisms were collected from the field and species identification is very difficult and not well-described in the literature. There were no other data available for similar organisms, so there is no question about combining values for a species mean value and the variability (as shown by the 95% confidence interval) is low, so it is reasonable to believe that the test organisms were all one species.

LC₅₀ values not calculated.

EC₅₀ calculated based on nominal concentrations.

Reliability points taken off for:

Documentation: Organism age (5), Conductivity (2), Hypothesis tests (8)

Acceptability: No standard method (5), Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Organisms randomized (1), Acclimation (1), Exposure type (2), Temperature variance (3), Conductivity (1), Random design (2), Appropriate statistical method (2), Hypothesis tests (3).

Pimephales promelas

Study: Kent SJ, Shillabeer N. 1997e. Lambda-cyhalothrin: Acute toxicity to fathead minnow (*Pimephales promelas*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Kent & Shillabeer 1997e	P. promelas
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Cypriniformes	
Family	Cyprinidae	
Genus	Pimephales	
Species	promelas	
Family in North America?	Yes	
Age/size at start of test/growth	NR	EPA
phase	>31 weeks old	recommendation
	mean control weight	0.5-5 g, but smaller
	and length 0.37 g and	fish should be more
	28 mm at end of test.	sensitive
Source of organisms	Continuous lab culture	Brixham
		Environmental
		Laboratory
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	Reared in same
free?		conditions
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	25 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
рН	7. 23-7.60	
Hardness	39.3-44.6 mg/L	
Alkalinity	22.7 mg/L	

Reference	Kent & Shillabeer 1997e	P. promelas
Parameter	Value	Comment
Conductivity	222-229 μS/cm	
Dissolved Oxygen	7.6-8.0 mg/L	
Feeding	None during test	
Purity of test substance	87.7%	
Concentrations measured?	Yes	
Measured is what % of nominal?	38-68%	
Chemical method documented?	Yes, GC-ECD	
Concentration of carrier (if any) in	0.01% dimethylformamide	
test solutions		
Concentration 1 Nom/Meas (µg/L)	0.060/0.025	1 rep, 20 org/rep
Concentration 2 Nom/Meas (µg/L)	0.125/0.082	1 rep, 20 org/rep
Concentration 3 Nom/Meas (µg/L)	0.25/0.17	1 rep, 20 org/rep
Concentration 4 Nom/Meas (µg/L)	0.50/0.34	1 rep, 20 org/rep
Concentration 5 Nom/Meas (µg/L)	1.0/0.38	1 rep, 20 org/rep
Concentration 6 Nom/Meas (µg/L)	2.0/1.3	1 rep, 20 org/rep
Control	Solvent and Dilution water	1 rep, 20 org/rep
LC ₅₀ (95% confidence interval)	24 h: 0.89 (0.73-1.1)	Method: Moving
$(\mu g/L)$	48 h: 0.89 (0.73-1.1)	average angle (24,
	72 h: 0.70 (0.38-1.3)	48 h); Binomial (72,
	96 h: 0.70 (0.38-1.3)	96 h)

LC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Photoperiod (3), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Pimephales promelas

Study: Tapp JF, Maddock BG, Harland BJ, Stembridge HM, Gillings E. 1990. Lambda-cyhalothrin (Karate PP321): Determination of chronic toxicity to fathead minnow (*Pimephales promelas*) full lifecycle. ICI Agrochemicals. Imperial Chemical Industries PLC, Brixham Laboratory, Brixham UK. MRID 41519001.

RelevanceReliabilityScore: 100Score: 93.5Rating: RRating: R

Reference	Tapp <i>et al.</i> 1990	P. promelas
Parameter	Value	Comment
Test method cited	EPA 1986 EPA 540/9-86-137	Some deviations from method, but determined to be scientifically sound
Phylum	Chordata	
Class	Osteichthyes	
Order	Cypriniformes	
Family	Cyprinidae	
Genus	Pimephales	
Species	promelas	Rafinesque
Family in North America?	Yes	
Age/size at start of test/growth phase	F ₀ : eggs when first exposed (chronic), larvae (acute) F ₁ : larvae when first exposed	
Source of organisms	Lab culture	Sea Plantation Eng. Tech. Salem, MA
Have organisms been exposed to contaminants?	No	
Animals acclimated and disease-free?	Yes	
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	300 d	
Data for multiple times?	Yes	96 hr, 7, 14, 21, 28, 42, 56 d
Effect 1	Hatching	
Control response 1	F0: 87.3% F1: 96.1%	Solvent and Dil water control pooled be no sig differences found
Effect 2	Survival	

Reference	Tapp et al. 1990	P. promelas			
Parameter	Value	Comment			
Control response 2	F ₀ : 96 hr 100%, 28d 93%, 56d 91%, 300 d 81.3% F ₁ : 56 d 79.6%	Solvent and Dil water control pooled be no sig differences found			
Effect 3	Length				
Control response 3	F ₀ : 20.95 mm (28 d), 31.4 mm (56 d), 52.5 mm (300 d) F ₁ : 25.0 mm (56 d)	Solvent and Dil water control pooled be no sig differences found			
Effect 4	Weight				
Control response 4	F ₀ : 3135.6 g (300 d) F ₁ : 240.4 g (56 d)	Solvent and Dil water control pooled be no sig differences found			
Effect 5	Egg production				
Control response 5	F ₀ : 74 eggs/batch 5918.8 total eggs produced	Solvent and Dil water control pooled be no sig differences found			
Temperature	$25 \pm 1^{\circ}\text{C}$				
Test type	FT				
Photoperiod/light intensity	16L:8D, 1100-1806 lux				
Dilution water	Filtered and dechlorinated tap water				
рН	6.09-8.36 (mean 7.2)				
Hardness	45.1 mg/L as CaCO ₃ (mean)	Range 32.6-57.0			
Alkalinity	26.1 mg/L as CaCO ₃ (mean)	Range 18.6-33.9			
Conductivity	100-160 uS/cm (mean 125 uS/cm)				
Dissolved Oxygen	7.06 mg/L (mean)	Not aerated			
Feeding	Yes, differed as fish aged	Not fed w/in 24 hr of weighing			
Purity of test substance	97%				
Concentrations measured?	Yes				
Measured is what % of nominal?	65-72% total cyhalothrin 50-56% l-cyhalothrin				
Chemical method documented?	Yes, LSC	GC/MS confirmation			
Concentration of carrier (if any) in test solutions	0.00125% 12.5 uL/L	Triethylene glycol			
Concentration 1 Nom/Meas (µg/L)	0.03/0.019/0.015	Duplicates and 40 eggs/rep, then 25 larvae/rep, then 4 mating pairs/rep			

Reference	Tapp <i>et al.</i> 1990	P. promelas
Parameter	Value	Comment
Concentration 2 Nom/Meas (µg/L)	0.06/0.040/0.031	Same as above
Concentration 3 Nom/Meas (µg/L)	0.12/0.081/0/062	Same as above
Concentration 4 Nom/Meas (µg/L)	0.25/0.180/0.139	Same as above
Concentration 5 Nom/Meas (µg/L)	0.50/0.354/0.273	Same as above
Concentration 6 Nom/Meas (µg/L)	1.0 – 96 hr test only	Same as above
Control	Dilution water and	Same as above
Control	solvent	Same as above
LC ₅₀ (95% confidence interval)	F ₀ 72 hr: 0.407 (0.316-	Method: Moving
(μg/L)	0.675)	average
(1-8/2)	F ₀ 96 hr: 0.360 (0.252-	
	0.765)	
	F ₀ 28 d: 0.114 (0.101-	
	0.130)	
	F ₀ 56 d: 0.108 (0.095-	
	0.124)	
	F ₁ 56 d: 0.059 (0.052-	
	0.067)	
NOEC (µg/L)	<u>F₀ generation</u>	Method: exact 2x2
	Hatch: ≥ 0.273	contingency table test
	28 d Survival: 0.062	(Hatch & % survival),
	56 d Survival: 0.062	Dunnett's t-tests, t-
	28 d Length: ≥ 0.139	tests, 1 way and 2 way
	56 d Length: ≥ 0.139	ANOVA (length &
	150-300 d Survival: <u>></u> 0.139	weight), t-tests & 1 way ANOVA (egg
	300 d Length: > 0.139	production)
	300 d Length: ≥ 0.139 300 d Weight: ≥ 0.139	p: 0.05
	Egg production: 0.062	MSD: NR
	255 production: 0.002	WISD, THE
	F ₁ generation	
	3-5 d Hatch: 0.062	
	56 d Survival: 0.031	
	56 d Length: \geq 0.139	
	56 d Weight: \ge 0.139	
	Overall: 0.031	
LOEC (µg/L)	<u>F₀ generation</u>	Same as above
	Hatch: > 0.273	
	28 d Survival: 0.139	
	56 d Survival: 0.139	
	28 d Length: > 0.139	
	56 d Length: > 0.139	
	150-300 d Survival: >	
	0.139	

Reference	Tapp <i>et al.</i> 1990	P. promelas
Parameter	Value	Comment
	300 d Length: > 0.139	
	300 d Weight: > 0.139	
	Egg production: 0.139	
	F ₁ generation	
	3-5 d Hatch: 0.139	
	56 d Survival: 0.062	
	56 d Length: > 0.139	
	56 d Weight: > 0.139	
	Overall: 0.062	
MATC (GeoMean NOEC,LOEC)	0.044 ug/L (>0.031,	
	<0.062)	
% control at NOEC	NR	
% of control LOEC	NR	

- □ All toxicity values calculated based on measured concentrations corrected mean measured concentrations of lambda-cyhalothrin (accounts for isomerization)
- \Box The 72/96 hr LC₅₀ test was run as a separate test but with the same organisms as from the F₀ tests

Bioconcentration factors (BCF) F₀ adults: 4982 (SD=1233) μg/L F₁ eggs: 1311 (SD=130) μg/L F₁ larvae: 4299 (SD=806) μg/L

Reliability points taken off for:

<u>Documentation:</u> Statistical significance (2), Minimum significant difference (2), % control at NOEC/LOEC (2)

<u>Acceptability:</u> Measured conc w/in 20% nominal (4), Random design (2), Minimum significant difference (1)

Poecilia reticulata

Study: Kent SJ, Shillabeer N. 1997b. Lambda-cyhalothrin: Acute toxicity to the guppy (*Poecilia reticulata*). ZENECA Agrochemicals. DPR study number 50907-085.

RelevanceReliabilityScore: 100Score: 84Rating: RRating: R

Reference	Kent & Shillabeer 1997b	P. reticulata
Parameter	Value	Comment
Test method cited	US EPA, OECD	
Phylum	Chordata	
Class	Osteichthyes	
Order	Cyprinodontiformes	
Family	Poeciliidae	
Genus	Poecilia	
Species	reticulata	
Family in North America?	Yes	
Age/size at start of test/growth	NR	
phase	> >17 d old	
	mean control weight	
	and length 2.15 g and	
	53 mm at end of test.	
Source of organisms	Lab culture	London Aquatic
		Co., UK
Have organisms been exposed to	No	
contaminants?		
Animals acclimated and disease-	Yes	17 d acclimation in
free?		facility
Animals randomized?	Yes	
Test vessels randomized?	NR	
Test duration	96 h	
Data for multiple times?	Yes; 24, 48, 72 h	
Effect 1	Mortality	
Control response 1	0%	
Temperature	12 ± 1 °C	
Test type	Flow-through	
Photoperiod/light intensity	NR	
Dilution water	Dechlorinated tap water	
рН	7.04-7.39	
Hardness	43.3-46.3 mg/L	
Alkalinity	25 mg/L	
Conductivity	212-218 μS/cm	

Reference	Kent & Shillabeer 1997b	P. reticulata		
Parameter	Value	Comment		
Dissolved Oxygen	9.2-10.4 mg/L, > 87% sat			
Feeding	None during test			
Purity of test substance	87.7%			
Concentrations measured?	Yes			
Measured is what % of nominal?	43-58%			
Chemical method documented?	Yes, GC-ECD			
Concentration of carrier (if any) in	0.01% dimethylformamide			
test solutions				
Concentration 1 Nom/Meas (µg/L)	0.030/0.017	1 rep, 20 org/rep		
Concentration 2 Nom/Meas (µg/L)	0.060/0.026	1 rep, 20 org/rep		
Concentration 3 Nom/Meas (µg/L)	0.12/0.056	1 rep, 20 org/rep		
Concentration 4 Nom/Meas (µg/L)	0.24/0.11	1 rep, 20 org/rep		
Concentration 5 Nom/Meas (µg/L)	0.48/0.28	1 rep, 20 org/rep		
Concentration 6 Nom/Meas (µg/L)	0.96/0.48	1 rep, 20 org/rep		
Control	Solvent and Dilution water	1 rep, 20 org/rep		
LC ₅₀ (95% confidence interval)	24 h: 0.18 (0.11-0.28)	Method: Binomial		
(µg/L)	48 h: 0.078 (0.056-0.11)			
	72 h: 0.078 (0.056-0.11)			
	96 h: 0.078 (0.056-0.11)			

LC₅₀ calculated based on measured concentrations.

Reliability points taken off for:

<u>Documentation:</u> Organism age (5), Photoperiod (3), Hypothesis tests (8)

Acceptability: Measured concentrations w/in 20% of nominal (4), Appropriate organism age (3), Photoperiod (2), Random design (2), Adequate replication (2), Hypothesis tests (3).

Procambarus clarkii

Study: Barbee GC, Stout MJ. 2009. Comparative acute toxicity of neonicotinoid and pyrethroid insecticides to non-target crayfish (*Procambarus clarkii*) associated with rice-crayfish crop rotations. Pesticide Management and Science, 65:1250-1256

RelevanceReliabilityScore: 100Score: 83.5Rating: RRating: R

Reference	Barbee & Stout 2009	P. clarkii		
Parameter	Value	Comment		
Test method cited	ASTM static renewal method	2002		
Phylum	Arthropoda			
Class	Malacostraca			
Order	Decapoda			
Family	Cambaridae			
Genus	Procambarus			
Species	clarkii			
Family in North America?	Yes			
Age/size at start of test/growth	3 months			
phase	6.7 cm long, 9.0 g			
Source of organisms	Outdoor university culture			
	canals			
Have organisms been exposed to	No			
contaminants?				
Animals acclimated and disease-	Yes			
free?				
Animals randomized?	Yes			
Test vessels randomized?	NR			
Test duration	96 h			
Data for multiple times?	No			
Effect 1	Mortality			
Control response 1	0 %			
Temperature	21.7 ± 1.5 °C			
Test type	Static renewal			
Photoperiod/light intensity	16 L: 8 D			
Dilution water	Filtered tap water			
pH	7.3			
Hardness	268 mg/L			
Alkalinity	214 mg/L			
Conductivity	NR			
Dissolved Oxygen	> 60 % saturation			
Feeding	None during test			

Reference	Barbee & Stout 2009	P. clarkii		
Parameter	Value	Comment		
Purity of test substance	99.1%			
Concentrations measured?	No			
Measured is what % of nominal?	n/a			
Chemical method documented?	No			
Concentration of carrier (if any) in	% NR, acetone			
test solutions				
Concentration 1 Nom (µg/L)	0.1	2 reps, 5 orgs/rep		
Concentration 2 Nom (µg/L)	0.2	2 reps, 5 orgs/rep		
Concentration 3 Nom (µg/L)	0.3	2 reps, 5 orgs/rep		
Concentration 4 Nom (µg/L)	0.6	2 reps, 5 orgs/rep		
Concentration 5 Nom (µg/L)	1.0	2 reps, 5 orgs/rep		
Control	Solvent and dilution water	2 reps, 5 orgs/rep		
LC ₅₀ (95% confidence limits)	0.16 (0.06-0.27) μg/L	Method: Probit		

LC50 calculated based on nominal concentrations.

Reliability points taken off for:

<u>Documentation:</u> Analytical method (4), Measured concentrations (3), Conductivity (2), Hypothesis tests (8)

<u>Acceptability:</u> Measured concentrations w/in 20% of nominals (4), Carrier solvent (4), Conductivity (1), Random design (2), Adequate replication (2), Hypothesis tests (3)

Various species (16)

Study: Schroer AFW, Belgers JDM, Brock TCM, Matser AM, Maund SJ, Vann den Brink PJ. 2004. Comparison of Laboratory Single Species and Field Population-Level Effects of the Pyrethroid Insecticide l-cyhalothrin on Freshwater Invertebrates. Arch Environ Contam Toxicol, 46: 324-335.

Relevance

Score: 67.5 (No standard method, low chemical purity, control response not acceptable)

Rating: N

Appendix B

Fit test calculations

11	Omit on	e											
all LC50s	1	2	3	4	5	6	7	8	9	10	11	12	13
0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023
0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028	0.0028
0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059	0.0059
0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130	0.0130
0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260	0.0260
0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300	0.0300
0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380	0.0380
0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	
0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780		0.0780
0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492		0.1492	0.1492
0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600		0.1600	0.1600	0.1600
0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600		0.1600	0.1600	0.1600	0.1600
0.2000	0.2000	0.2000	0.2000	0.2000	0.2000	0.2000	0.2000		0.2000	0.2000	0.2000	0.2000	0.2000
0.2717	0.2717	0.2717	0.2717	0.2717	0.2717	0.2717		0.2717	0.2717	0.2717	0.2717	0.2717	0.2717
0.3000	0.3000	0.3000	0.3000	0.3000	0.3000		0.3000	0.3000	0.3000	0.3000	0.3000	0.3000	0.3000
0.4000	0.4000	0.4000	0.4000	0.4000		0.4000	0.4000	0.4000	0.4000	0.4000	0.4000	0.4000	0.4000
0.5020	0.5020	0.5020	0.5020		0.5020	0.5020	0.5020	0.5020	0.5020	0.5020	0.5020	0.5020	0.5020
0.6400	0.6400	0.6400		0.6400	0.6400	0.6400	0.6400	0.6400	0.6400	0.6400	0.6400	0.6400	0.6400
2.3000	2.3000		2.3000	2.3000	2.3000	2.3000	2.3000	2.3000	2.3000	2.3000	2.3000	2.3000	2.3000
3.3000		3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000	3.3000

	Omit one continued from above												
all LC 50s	14	15	16	17	18	19	20						
0.0023	0.0023	0.0023	0.0023	0.0023	0.0023	0.0023							
0.0028	0.0028	0.0028	0.0028	0.0028	0.0028		0.0028						
0.0059	0.0059	0.0059	0.0059	0.0059		0.0059	0.0059						
0.0130	0.0130	0.0130	0.0130		0.0130	0.0130	0.0130						
0.0260	0.0260	0.0260		0.0260	0.0260	0.0260	0.0260						
0.0300	0.0300		0.0300	0.0300	0.0300	0.0300	0.0300						
0.0380		0.0380	0.0380	0.0380	0.0380	0.0380	0.0380						
0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470						
0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780	0.0780						
0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492	0.1492						
0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600						
0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600	0.1600						
0.2000	0.2000	0.2000	0.2000	0.2000	0.2000	0.2000	0.2000						
0.2717	0.2717	0.2717	0.2717	0.2717	0.2717	0.2717	0.2717						
0.3000	0.3000	0.3000	0.3000	0.3000	0.3000	0.3000	0.3000						
0.4000	0.4000	0.4000	0.4000	0.4000	0.4000	0.4000	0.4000						

0.5020

0.6400

2.3000

3.3000

0.5020

0.6400

2.3000

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0.6400

2.3000

3.3000

Omitted point, xi:	0.0023	0.0028	0.0059	0.0130	0.0260	0.0300	0.038	0.0470	0.0780	0.1492	0.1600	0.1600	0.2000
median 5th percentile	0.0045	0.0042	0.0033	0.0026	0.0022	0.0022	0.002	0.002	0.002	0.0022	0.0023	0.0023	0.0023
Burr III													
percentile	97.04	96.29	92.55	86.43	78.57	76.58	72.98	69.37	59.13	43.12	41.37	41.37	35.95
F-i(xi)	0.9704	0.9629	0.9255	0.8643	0.7857	0.7658	0.729		0.5913	0.4312	0.4137	0.4137	0.3595
1-F(xi)	0.0296	0.0371	0.0745	0.1357	0.7637	0.2342	0.270		0.4087	0.5688	0.5863	0.5863	0.6405
1-r(xi)	0.0290	0.0371	0.0743	0.1337	0.2143	0.2342	0.270	2 0.3003	0.4007	0.5000	0.5605	0.5605	0.0403
Min of F-i(xi) or 1-F(xi)	0.0296	0.0371	0.0745	0.1357	0.2143	0.2342	0.2702		0.4087	0.4312	0.4137	0.4137	0.3595
$p_i = 2(min)$	0.0592	0.0742	0.149	0.2714	0.4286	0.4684	0.540	4 0.6126	0.8174	0.8624	0.8274	0.8274	0.719
Continued from above													
Omitted point, xi:	0.2717	0.3	8000	0.4000	0.5020	0.64	00 2	2.3000	3.3	_			
median 5th percentile	0.002	4 0	.0025	0.0025	0.0025	0.00	25 0	.0021	0.0019				
Burr III													
percentile	29.1	.6	27.13	21.74	17.97	14	.38	2.38	0.97				
F-i(xi)	0.2916	5 0.2	2713	0.2174	0.1797	0.14	38	0.0238	0.0097				
1-F(xi)	0.7084	0.7	287	0.7826	0.8203	0.85	62	0.9762	0.9903				
. /													
Min of F-i(xi) or 1-F(xi)	0.2916	5 02	2713	0.2174	0.1797	0.14	38 1	0.0238	0.0097				
. , , , , ,	0.5832		5426	0.4348		0.28		0.0476	0.0097				
$p_i = 2(min)$	0.3832	2 0.5	420	0.4348	0.3594	0.28	/0 (J.U4 / O	0.0194				

	-	Fisher test statis	tic
p _i	ln(p _i)	-2*Sum of ln (pi)	X^2_{2n}
0.0592	-2.8268	47.2916	0.1994
0.0742	-2.6010		
0.1490	-1.9038		
0.2714	-1.3042		
0.4286	-0.8472		
0.4684	-0.7584		
0.5404	-0.6154		
0.6126	-0.4900		
0.8174	-0.2016		
0.8624	-0.1480		
0.8274	-0.1895		
0.8274	-0.1895		
0.719	-0.3299		
0.5832	-0.5392		
0.5426	-0.6114		
0.4348	-0.8329		
0.3594	-1.0233		
0.2876	-1.2462		
0.0476	-3.0449		
0.0194	-3.9425		